



## The Efficacy of Synbiotic Application in Broiler Chicken Diets, Alone or in Combination with Antibiotic Growth Promoters on Zootechnical Parameters

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### ABSTRACT

In recent years, probiotics and synbiotics have gained considerable interest in poultry feeding as an alternative to antibiotics due to antibiotic resistance concerns. The objective of this dual study was to evaluate the efficacy of synbiotic supplementation alone or in combination with different Antibiotic Growth Promoters (AGPs), compared to the untreated control group of broiler chickens production performance. In the first experiment, a total of 1260 one-day-old male Ross 308 broiler chickens were randomly assigned to 7 diet treatments, with 6 replicates per diet treatment and 30 birds per replicate over a 42-day period. The diet treatments included a control diet based on corn-soybean without additives (T1), and the diet treatment with bacitracin (BMD 100 ppm, T2), colistin (10 ppm, T3), synbiotic (PoultryStar me, 0.5 kg/t, T4), a combination of synbiotic (0.5 kg/t) and bacitracin (60 ppm, T5), synbiotic (0.5 kg/t) and colistin (5 ppm, T6), synbiotic (0.5 kg/t), bacitracin (60 ppm), and colistin (5 ppm, T7). During the critical period of rearing from hatch to day 10, the synbiotic supplementation resulted in a significantly higher body weight gain than its combination with bacitracin. No other dietary treatment showed a remarkable improvement in the body weight gain, feed intake, or feed conversion ratio, compared to the only synbiotic application (T4) during the entire trial period. The tendency towards an improved feed conversion ratio was observed during the use of synbiotic (T4, 1.87), compared to the control group (T1, 1.93) during the entire trial period. Compared with the control group (T1, 2.78%), broiler mortality was also lower in the synbiotic group (T4, 1.11%). In the second experiment, a total of 1500 one-day-old male Ross 308 broiler chickens were randomly assigned to 4 diet treatments; with 15 replicates per diet treatment, and 25 birds per replicate over a 42-day period. The dietary treatments included a control group diet based on corn-soybean without additives (T1), and the treatment diets with bacitracin (BMD 1000 ppm, T2), synbiotic (PoultryStar me, 0.5 kg/t, T3), and a combination of synbiotic (0.5 kg/t) plus bacitracin (BMD 1000 ppm T4). Birds fed antibiotic or synbiotic alone or in a combination had numerically a higher body weight and an average daily gain than the control group. There was a tendency of improvement in the feed conversion ratio during the age of 1-24 days, and throughout the experimental period. The evaluated synbiotic could serve as an effective alternative to AGPs, such as bacitracin and colistin in broiler chicken diets, especially during the first crucial period. The synbiotic can serve this purpose without combining it with AGPs, such as colistin or bacitracin.

**Keywords:** Antibiotic growth promoter, Broilers, Performance, Synbiotic

### INTRODUCTION

In view of the apprehensions of antibiotic resistance, probiotics have gained considerable interest in the poultry industry as alternatives to antibiotics (Gustafson and Bowen, 1997). Presently, this class of feed additives is largely used as an alternative to Antibiotic Growth Promoters (AGP) in poultry feeding. The main impetus that has catalyzed the use of these probiotic feed additives is the worldwide ban on the use of AGPs in the diets of food animals. The alternatives to AGPs should ideally

possess the same beneficial effects as AGPs do possess when they are supplemented in the diet of food animals.

Despite the incredulous mechanism of action of the feed antibiotics (Huyghebaert et al., 2011), it is generally believed that the AGPs depict some antibacterial activities, which reduces the incidence and severity of subclinical infections, and decreases the microbial consumption of nutrients, thus improving the absorption of nutrients (Snyder and Wostmann, 1987; Brennan et al., 2003). The subsequent effect of all these activities leads to a better

performance of the animal. The foundation of this explanation lies in the fact that AGPs do not exert growth-promoting effects in germ-free animals. The prevailing practice of the industry to feed livestock with sub-therapeutic doses of antibiotics is unlikely to have a growth inhibitory effect on the resident bacteria (Niewold, 2007). However, when antibiotics were added to the broiler diets at the levels below minimum inhibitory concentration, a clear shift in the intestinal microbiota was observed which at least partly explains the effects of AGPs (Pedroso *et al.*, 2006; Wise and Siragusa, 2007).

Shifts in intestinal microbiota likewise affected the intestinal wall morphology and induced immune reactions which may promote the host animals' growth by affecting their energy expenses (Teirlynck *et al.*, 2009). Thus, AGP-alternatives such as probiotics as the hypothetical AGP mode of action should also have modulatory effects on intestinal microbiota and immune system. Probiotics are live microorganisms that should be viable when they are administered in the livestock diets; in order to exert their beneficial effects on an improved intestinal function, intestinal microbiota balance, host immune responses, and the overall host health (FAO and WHO joint working group, 2002).

Dietary probiotics contribute to establish and maintain a beneficial intestinal microbiota, which may enhance the colonization resistance to pathogens, and strengthen the immune responses, leading to an improved growth performance (Dhama *et al.*, 2011; Yang *et al.*, 2012; Mountzouris, 2014; Mountzouris *et al.*, 2015). The path of considering the gastrointestinal tract of food animals as the real complexity of anatomical system playing digestive, absorptive, metabolic, immunological, and endocrinological roles has progressed a lot in the last three decades (Oviedo-Rondon, 2019), the reason why the asseveration gut health became collectively important for the researchers and the livestock industry (Kogut *et al.* 2017).

The supplementation of probiotics and prebiotics has shown promising results in controlling bacterial infections in poultry by positively influencing the gut microbiota (Mead, 2000). Probiotics competitively excluded pathogenic microbes (Nava *et al.*, 2005) and can be effective by stimulating the immune responses (Koenen *et al.*, 2004), producing antibacterial substances, and stimulating digestive enzymes secretion (Saarela *et al.*, 2000). Synergistic effects could be achieved through so-called synbiotics, a combination of probiotics and prebiotics (Roberfroid, 1998). The combined supplementation of poultry diets with probiotics and

prebiotics (synbiotic) has been reported to be more effective than a single supplementation and in some cases even congruous with antibiotic treatments as reported in several studies and reviews (Gaggia *et al.*, 2010; Gadde *et al.*, 2017; Tayeri *et al.*, 2018). Improvements in feed efficiency in broiler chickens as a result of synbiotic supplementation have been attributed to their potential modulatory effect on gastro-intestinal microbial colonization (Brugaletta *et al.*, 2020). Prebiotics are indigestible carbohydrates supplemented frequently in combination with probiotics, which could stimulate the growth of useful bacteria in the intestines of the host (Lee *et al.*, 2016). Prebiotic supplementation was shown to mimic the attachment sites of the pathogens, decreasing the adherence of pathogenic bacteria to the intestinal wall, and increasing specific beneficial bacteria (Ija and Tivey, 1998). Therefore, it draws a great interest to evaluate the effects of a synbiotic on the broiler chicken's performance.

The synbiotic product (PoultryStar me, Biomin Holding GmbH, Austria) evaluated in previous studies contained probiotic bacterial strains of *Enterococcus*, *Bifidobacterium*, *Pedicoccus*, and *Lactobacillus* species and a prebiotic fructooligosaccharide (Babazadeh *et al.*, 2011).

Given the growth-promoting and immune-modulatory roles of AGPs (Niewold, 2007; Kogut and Swaggerty, 2012; Mountzouris, 2014), the performance response of the broilers to synbiotic products, when experimentally supplemented with AGPs in different combinations had been largely unknown. It was not clear whether there were additive effects due to the combination of AGPs and synbiotics.

The aim of these two experimental trials was therefore to evaluate the effect of dietary inclusion of a specific multi-species poultry synbiotic product alone or in different combinations with Bacitracin and/or Colistin, which are used as AGPs on the performance parameters in broiler chickens.

## MATERIALS AND METHODS

### Ethical approval

All procedures were performed in compliance with relevant laws and institutional guidelines. All animal experiments comply with the ARRIVE guidelines and were carried out in accordance with the U.K. Animals (Scientific Procedures) Act, 1986 and associated guidelines, EU Directive 2010/63/EU for animal experiments.

## First experiment

### *Animals and bird husbandry*

A 42-day broiler feeding trial was conducted at the Poultry Research and Development Center of Kasetsart University in Kamphangsae, Nakhon Pathom, Thailand with a total of 1261 day-old male Ross 308 broiler chickens (with an average body weight of 45 grams at 6-hours post-hatching), according to the prevailing institutional ethical norms. The chickens were weighed individually and assigned to seven treatment groups, each comprising of 6 replicates (n = 30 chickens on the first day). All chickens were raised in floor pens with rice husk as the litter material. Each compartment was equipped with manual feeders and bell-shaped drinkers without nipples. Feed (starter mash from day 1 to 10, grower mash from day 11 to 24, finisher mash 1 from day 25 to 35, and finisher mash 2 from day 36 to 42), and water were offered *ad libitum*. The lighting program was 23-hours light, and 1-hour dark period during the study. The chickens were housed in the evaporative cooling system during the experimental period. The chickens were vaccinated against Newcastle Disease (ND live B1) and Infectious Bronchitis on day 7, Infectious Bursal disease on day 14, and against Newcastle disease (La Sota strain) and Infectious Bronchitis on day 21 again. The temperature was maintained around 32 to 34°C for the first week, and then reduced weekly from 34°C to 25°C. The clinical observations regarding the animal health status, as well as the temperature, humidity, ventilation, and lighting of the trial house, were recorded daily during the experimental period.

### *Experimental diets and treatments*

The trial chickens were randomly assigned to 7 dietary treatments. Each treatment consisted of 6 replications with 30 chickens per replication using a completely randomized design to minimize the effects of group compartments.

All experimental diets were based on the corn-soybean meal. The dietary treatments are presented in tables 1 and 2.

The ingredients and the chemical composition of the experimental diets are presented in table 3, and the nutrient composition of the experimental diets (proximate analysis) is presented in table 4. The synbiotic product used in the present study was obtained from Biomin Holding GmbH, Getzersdorf, Austria, and was included in the diet according to the manufacturer's recommendation. The multi-species product synbiotic (PoultryStar<sup>®</sup> me) contained probiotic bacterial strains of *Enterococcus*, *Pediococcus*, *Bifidobacterium*, and *Lactobacillus* species as well as a prebiotic fructooligosaccharide.

All the diets were analyzed (AOAC, 2016) for Dry Matter (DM, method 934.01), crude protein (method 988.05), Crude Fiber (method 962.09, CF, Foss Fiber Cap 2021 Fiber Analysis System, Foss Analytical, Hilleroed, Denmark), and crude fat (petroleum ether extraction; method 920.39). Feed samples of each experimental diet prepared for the trial were collected per phase and group immediately after blending and mixing.

### *Performance parameters measurement*

Chicken live weight was recorded individually for each pen on the days 0, 10, twenty-four, and thirty-five, and per each group on the day forty-two. Body Weight Gain (BWG) was calculated per each group. Furthermore, Feed Intake (FI) was measured for the respective periods in combination with body weight measurements. Hence, the average of FI was determined for the respective periods per each group. Feed Conversion Ratio (FCR) for the respective periods was calculated for each group as the mortality-adjusted ratio between FI and BWG.

## Second experiment

### *Animals and birds' husbandry*

One thousand five hundred, one day old, male Ross 308 broiler chicks were divided into 4 dietary treatment groups. Each treatment comprised of fifteen replications with twenty-five chickens per replication, and the housing conditions were identical as in the first experimental trial.

### *Experimental diets and treatments*

All diets were corn-soybean meal, formulated to meet the nutritional requirements recommended by Ross

308 nutrition specification guide as in the first experiment. The trial began when the birds were one-day-old, and it was finalized when they were forty-two days old. The chickens were divided into 4 dietary treatment groups. Each treatment consisted of fifteen replications with twenty-five birds per replication, followed by a fully randomized design to minimize the effects of group compartments. The dietary treatments are presented in table 2.

The ingredients and the chemical composition of the experimental diets are presented in table 5, and the nutritional composition of the experimental diets (proximate analysis) is presented in table 6. The synbiotic product used in the present study was similar to the first trial, obtained from Biomin Holding GmbH, Getzersdorf, Austria, and was included in the diet as recommended by the manufacturer. The multi-species product synbiotic (PoultryStar® me) contained probiotic bacterial strains of *Enterococcus*, *Pediococcus*, *Bifidobacterium*, and

*Lactobacillus* species, and a prebiotic fructooligosaccharide. All other details of the conditions and practices related to the preparations, mixing, and application procedures of the experimental diets were similar to those of the first trial.

**Measurement of performance parameters**

The procedures for measuring the performance parameters were the same as described for the first experiment.

**Statistical analysis**

The pens were the experimental units, and all data were pooled per pen, unless specified different and expressed as the mean, and pooled the Standard Error of Means (SEM). The data were subjected to a one-way analysis of variance (Statistical Package for Social Sciences, SPSS version 10.1) with the diets as the factor, and it was found to be significant. The means were separated by Duncan’s new multiple range test at  $p < 0.05$ .

**Table 1.** Description of the Dietary treatments applied to Ross 308 broiler chickens

Treatment groups	Description
T1= Negative control (NC)	No additives in feed
T2 = Positive control (PC) 1 (AGP1)	Bacitracin (100 ppm* active ingredient).
T3 = Positive control (PC) 2 (AGP2)	Colistin (10 ppm active ingredient)
T4 = Synbiotic	Synbiotic (PoultryStar me 0.5 kg/ton of feed)
T5 = Synbiotic + PC 1 (PS**+AGP1***)	Synbiotic 0.5 kg/ton feed + Bacitracin (60 ppm active ingredient)
T6 = Synbiotic + PC 2 (PS+AGP2****)	Synbiotic 0.5 kg/ton feed + Colistin (5 ppm active ingredient)
T7 = Synbiotic + PC 1 + PC2 (PS+AGPs)	Synbiotic 0.5 kg/ton feed + Bacitracin (60 ppm active ingredient) + Colistin (5 ppm active ingredient)

\*ppm: parts per million, \*\*PoultryStar me, \*\*\* Antibiotic Growth Promoter 1 (Bacitracin), \*\*\*\* Antibiotic Growth Promoter 2 (Colistin)

**Table 2.** Description of the Dietary treatments applied to Ross 308 broiler chickens

Treatment groups	Description
T1= Negative control (NC)	No additives in feed
T2 = Positive control (PC) 1 (AGP)	Bacitracin (BMD*** 10% 1000 ppm*).
T3 = Synbiotic	PoultryStar 0.5 kg/t**** of feed
T4 = Synbiotic with AGP**	Poultry Star 0.5 kg / t + Bacitracin (BMD 10% 1000 ppm)

\*ppm: parts per million, \*\*Antibiotic Growth Promoter Bacitracin, \*\*\* Bacitracin methylene di-salicylate, \*\*\*\* kilogram per ton

**Table 3.** Ingredient composition and calculated analysis of experimental diets fed to the Ross 308 broiler chickens during the 42-day trial in the facility of Kasetsart University.

Ingredients	Unit	Starter (Day 1-10)	Grower (Day 11-24)	Finisher 1 (Day 25-35)	Finisher 2 (Day 36-42)
Corn	%	53.40	57.17	61.69	61.69
Soybean meal (46 % CP)	%	30.78	25.94	19.77	19.77
Full fat soybean (35.5 % CP)	%	12.00	13.50	15.00	15.00
Rice bran oil	%	0.50	0.50	1.24	1.24
MDCP (16.9 % Ca, 21.6 % P)	%	0.52	0.33	0.09	0.09
Limestone (38.7 % Ca)	%	0.96	0.87	0.72	0.72
Salt	%	0.41	0.41	0.39	0.39
Sodium bicarbonate (27 % Na)	%	0.05	0.05	-	-
Choline chloride (60 %)	%	0.04	0.03	0.04	0.04
Premix	%	0.60	0.60	0.60	0.60
L-Lysine	%	0.28	0.22	0.19	0.19
DL-Methionine	%	0.26	0.22	0.20	0.20
L-Threonine	%	0.13	0.09	0.06	0.06
Salinomycin (66 ppm)	%	0.05	0.05	0.05	-
Lutanox	%	0.02	0.02	0.02	0.02
Phytase	%	0.01	0.01	0.01	0.01
Total	%	100.00	100.00	100.00	100.00
<b>Calculated analysis</b>					
ME for poultry	kcal/kg	3053	3100	3200	3200
Protein	%	23.00	21.50	19.50	19.50
Fat	%	5.02	5.41	6.54	6.54
Fiber	%	4.00	3.90	3.72	3.72
Digestible Lysine (Poultry)	%	1.28	1.15	1.02	1.02
Digestible Methionine (Poultry)	%	0.51	0.47	0.43	0.43
Digestible Threonine (Poultry)	%	0.88	0.79	0.70	0.70
Lysine	%	1.44	1.30	1.17	1.17
Methionine + Cysteine	%	0.92	0.87	0.80	0.80
Methionine	%	0.61	0.55	0.50	0.50
Threonine	%	0.97	0.88	0.78	0.78
Calcium	%	0.80	0.72	0.61	0.61
Total phosphorus	%	0.60	0.55	0.48	0.48
Avail. Phosphorus (poultry)	%	0.33	0.29	0.24	0.24
Choline	%	1700	1600	1500	1500
Sodium	%	0.19	0.19	0.17	0.17
Salt	%	0.45	0.45	0.42	0.42

ME = Metabolizable Energy, MDCP = Mono-Dicalcium Phosphate.

**Table 4.** Nutrient composition of experimental diets (proximate analysis) fed to the Ross 308 broiler chickens during the 42-day trial in the facility of Kasetsart University.

Item	Period			
	Starter	Grower	Finisher1	Finisher2
Protein (%)	22.31	20.54	18.25	18.68
Fiber (%)	4.39	4.66	4.08	3.87
Fat (%)	6.01	6.26	5.49	6.08
Ash (%)	4.6	4.69	3.87	3.84
Calcium (%)	0.79	0.8	0.63	0.57
Phosphorus (%)	0.41	0.43	0.34	0.33
GE (kcal/kg)	4.657.52	4.684.73	4.530.39	4.603.69

GE = Gross Energy

## RESULTS

### First experiment's results

All chickens were healthy during the experimental period, and there was no mortality during the most critical period from the hatch to day 10. The outcome depicted

that the synbiotic supplementation in the broiler diet resulted in a significantly higher Body Weight Gain (BWG) than its combination with bacitracin ( $p < 0.05$ ) during the hatch to day 10 (Table 7). Additionally, the treatment groups T3 (colistin alone) and T6 (colistin with synbiotic) resulted in a significantly better ( $p < 0.05$ ) BWG

during this period compared to the control group (Table 7). None of the other treatments improved BWG, FI, or FCR significantly compared to the only synbiotic application (T4) during the entire experimental period from the hatch to day 42 (Table 8). An improved FCR ( $p = 0.0756$ ) of 1.86 was observed in the bacitracin group (T2), 1.87 in the symbiotic group (T4), and 1.83 in the synbiotic-AGPs combination group (T7), respectively compared to the control group (T1, 1.93), and other treatment groups during the entire trial period (Table 8). No mortality was observed in the colistin-synbiotic combination group (T6) during the entire trial period. However, remarkably low mortality of 1.11% occurred in the bacitracin group (T2), the synbiotic group (T4), and the synbiotic-AGPs combination group (T7), respectively during the entire trial period compared to the control group (T1, 2.78%) and other treatment groups (Table 8).

### Second experiment's results

The birds were healthy throughout the entire experimental trial. The crude protein contents in the mixed feeds corresponded to the calculated values. The amount of crude fat, crude fiber, Calcium (Ca), and phosphorous (P) in the experimental diets also was confirmed well by the calculated values (Table 5). Although no significant differences between the dietary treatments regarding zootechnical parameters were observed, the birds fed only with AGP or synbiotic and AGP in combination with synbiotic had a numerically higher body weight and average daily BWG than the non-supplemented control groups ( $p = 0.2500$ ). This led to a tendency to improve FCR between the age of 1 to twenty-four days old, and throughout the experimental period of 1 to forty-two days (Tables 9 and 10).

**Table 5.** Ingredient composition and calculated analysis (%) of the second experimental diets fed to the Ross 308 broiler chickens during the 42-day trial in the facility of Kasetsart University.

Ingredient	Unit	Starter	Grower	Finisher
Corn	%	54.75	59.52	64.30
Soybean oil	%	1.92	1.72	1.50
Soybean Meal 48 %	%	30.65	24.48	18.25
Full fat Soybean	%	8.00	10.00	12.00
Calcium carbonate	%	1.45	1.33	1.22
MCP-22	%	1.79	1.60	1.44
Salt	%	0.36	0.36	0.36
DL-Methionine	%	0.34	0.30	0.26
L-Lysine	%	0.25	0.23	0.23
Threonine	%	0.09	0.07	0.04
Choline Chloride 60%	%	0.06	0.06	0.05
Antioxidant	%	0.01	0.01	0.01
Toxin Binder	%	0.15	0.15	0.15
Premix (vitamin + mineral)	%	0.18	0.18	0.18
Total	%	100.00	100.00	100.00
<b>Calculated analysis</b>				
ME for Poultry	kcal/kg	3100.00	3150.00	3200.00
Protein	%	23.00	21.00	19.00
Moisture	%	10.92	10.97	11.03
Fat	%	5.93	6.24	6.53
Fiber	%	3.15	3.20	3.26
Ash	%	5.80	5.31	4.86
Ca	%	0.96	0.87	0.79
Total P	%	0.77	0.71	0.65
P avail	%	0.48	0.44	0.40
Salt	%	0.36	0.35	0.35
Lysine	%	1.44	1.29	1.16
Methionine	%	0.67	0.61	0.55
Methionine + Cysteine	%	1.08	0.99	0.91
Threonine	%	0.97	0.88	0.78
Tryptophan	%	0.28	0.25	0.22
Arginine	%	1.54	1.39	1.24
Choline Chloride	mg/kg	1700.00	1600.00	1500.00

ME = Metabolizable Energy, MCP = Monocalcium Phosphate 22% feed grade



**Table 6.** Nutrient composition of the second experimental diets (proximate analysis) fed to the Ross 308 broiler chickens during the 42-day trial in the facility of Kasetsart University.

Nutrient (%)	Treatment 1	Treatment 2	Treatment 3	Treatment 4
<b>Starter</b>				
Moisture	11.60	11.29	11.48	11.34
Protein	21.93	21.48	21.93	22.31
Fat	5.98	5.70	5.48	5.52
Fiber	2.38	2.44	2.40	2.39
Ash	5.81	5.77	5.72	5.69
Calcium	1.01	0.97	1.00	1.00
Phosphorus	0.80	0.74	0.78	0.77
GE (kcal/kg)	4092.82	4106.91	4174.68	4187.77
<b>Grower</b>				
Moisture	11.44	11.29	10.76	11.18
Protein	19.77	19.78	20.10	19.72
Fat	5.96	6.08	6.13	6.01
Fiber	2.03	2.06	2.15	1.95
Ash	6.23	6.32	6.31	6.16
Calcium	0.89	0.91	0.88	0.92
Phosphorus	0.71	0.74	0.73	0.71
GE (kcal/kg)	4147.89	4180.21	4238.82	4230.72
<b>Finisher</b>				
Moisture	12.01	12.02	11.75	11.70
Protein	18.90	18.78	18.98	18.67
Fat	6.70	6.65	6.40	6.52
Fiber	2.06	2.22	2.02	2.12
Ash	4.82	4.83	4.89	4.94
Calcium	0.80	0.82	0.82	0.79
Phosphorus	0.63	0.63	0.64	0.61
GE (kcal/kg)	4373.33	4385.00	4253.81	4218.67

GE = Gross Energy, ME = Metabolizable Energy

## DISCUSSION

Presently, probiotics are largely used as alternatives to antibiotic growth promoters (AGP) in the modern poultry nutrition due to concerns of antibiotic resistance, and the ban imposed on the usage of AGPs in the diets of food animals. Beneficial effects of single or multi-species probiotics on the zootechnical performance of broiler chickens were increasingly documented in the scientific literature (Applegate et al., 2010; Fuentes et al., 2013; Zhang and Kim, 2014; Gadde et al., 2017; Tayeri et al., 2018). The data strongly suggested an improvement in health throughout the experimental period and the complete absence of mortality during the most critical period of day 0 to 10 (Table 7). This is in concordance with studies by Pelicano et al. (2004), and Takahashi et al. (2005), in which the use of different growth promoters in the early phase of rearing led to no differences in the viability and mortality rates of the broiler chickens. The present results indicated that the synbiotic supplementation in the diets of broiler chickens resulted in a significantly higher BWG than the combination with bacitracin ( $p < 0.05$ ) during the first days of the post-hatch

brooding period, considered the most critical phase of rearing from hatch to day 10 (Table 7). Probiotics are known to contribute towards the establishment and maintenance of a beneficial intestinal microbiota, which could enhance the colonization resistance to pathogens, and immune response improvements resulting in improved growth performance (Mountzouris, 2014; Mountzouris et al., 2015; Kogut et al. 2017; Baldwin et al., 2018; Oviedo-Rondon, 2019; Brugaletta et al., 2020).

The body weight gain was significantly better ( $p < 0.05$ ) in T3 (colistin alone) and T6 (colistin with synbiotic) treatment groups compared to the control groups during this critical period from hatch to day 10 of age (Table 7). Synergistic effects were observed by feeding synbiotics, which are a combination of probiotics and prebiotics (Roberfroid, 1998; Gaggia et al., 2010; Gadde et al., 2017; Tayeri et al., 2018). There was not any significant improvement in BWG, FI, or FCR in any other group compared to the only synbiotic application (T4) during the entire study period from the hatching day to the day forty-second (Table 8).

An improved FCR ( $p=0.0756$ ) of 1.86 was observed in the bacitracin group (T2), 1.87 in the synbiotic group

(T4), and 1.83 in the synbiotic-AGPs combination group (T7), respectively compared to the control group (T1, 1.93), and other treatment groups in the study (Table 8). No mortality was recorded in the colistin-synbiotic combination group (T6) during the entire trial period. In the bacitracin group (T2), the synbiotic group (T4), and the synbiotic-AGPs combination group (T7), however, a very low bird mortality rate of 1.11% occurred compared to the control group (T1, 2.78%), and other treatment groups (Table 8). Chickens in the second experimental trial were also healthy during the entire study. Although no significant differences among the dietary treatments regarding zootechnical parameters were observed, birds

fed with AGP or synbiotic alone, and their combination, had a numerically higher body weight and average daily BWG than that of the control groups ( $p=0.2500$ ). This improvement in the BWG in these treatment groups tended to improve FCR in the chickens aged 1 to twenty-four days, and 1 to forty-two days old throughout the experimental period (Tables 9 and 10). No significant differences in body weight, FI, FCR, and mortality among the synbiotic, colistin, and bacitracin groups alone or in combination with each other revealed that AGP could be replaced by synbiotics without loss of zootechnical performance.

**Table 7.** Effect of the combination of synbiotics with antibiotic growth promoters on the production performance of broiler chickens from day of hatch to day 10.

Treatment groups <sup>1</sup>	BWG	FI	FCR	Mortality (%)
	(g/bird)	(g/bird)		
T1	192.656 <sup>ab</sup>	263.472	1.36	0.00
T2	193.094 <sup>ab</sup>	272.611	1.41	0.00
T3	199.133 <sup>a</sup>	271.389	1.36	0.00
T4	198.461 <sup>a</sup>	270.217	1.36	0.00
T5	186.050 <sup>b</sup>	272.361	1.46	0.00
T6	199.678 <sup>a</sup>	267.611	1.34	0.00
T7	193.189 <sup>ab</sup>	265.583	1.37	0.00
p -value	0.0337	0.6047	0.1492	0.00
SEM	1.2663	1.5107	0.0129	0.00

<sup>a,b</sup> Means with dissimilar letters in a column varied significantly ( $p < 0.05$ ) <sup>1</sup> T1= No additives in feed, T2 = Bacitracin, T3 = Colistin, T4 = Synbiotic (PoultryStar<sup>®</sup> me), T5 = Synbiotic + Bacitracin, T6 = Synbiotic + Colistin, T7 = Synbiotic + Bacitracin + Colistin. BWG = Body Weight Gain, FI = Feed Intake, FCR = Feed Conversion Ratio, SEM = Standard Error of Means

**Table 8.** Effect of the combination of synbiotics with antibiotic growth promoters on the production performance of broiler chickens from the first day to day 42.

Treatment groups <sup>1</sup>	BWG	FI	FCR	Mortality (%)
	(g/bird)	(g/bird)		
T1	2872.50	5547.01	1.93	2.78
T2	2921.17	5435.59	1.86	1.11
T3	2871.02	5540.05	1.93	1.67
T4	2923.90	5479.02	1.87	1.11
T5	2883.07	5412.70	1.88	1.67
T6	2893.12	5444.65	1.88	0.00
T7	2972.42	5453.08	1.83	1.11
p-value	0.7736	0.6241	0.0756	0.4758
SEM	18.1249	22.4774	0.0096	2.2160

<sup>1</sup> T1= No additives in feed, T2 = Bacitracin, T3 = Colistin, T4 = Synbiotic (PoultryStar<sup>®</sup> me), T5 = Synbiotic + Bacitracin, T6 = Synbiotic + Colistin, T7 = Synbiotic + Bacitracin + Colistin. BWG = Body Weight Gain, FI = Feed Intake, FCR = Feed Conversion Ratio, SEM = Standard Error of Means.



**Table 9.** Effect of dietary treatments on growth performance of Ross 308 broiler chickens from day 1 to day 24, fed in the facility of Kasetsart University.

Treatment	Feed intake (g)	Body weight (g)	ADG (g/bird/day)	FCR	% Livability
NC	1720.573	1013.843	42.243	1.698	99.733
AGP	1717.280	1021.277	42.553	1.682	100.000
PS	1721.093	1021.189	42.550	1.687	99.733
PS + AGP	1721.107	1020.827	42.534	1.686	99.733
p-value	0.9818	0.8752	0.8752	0.6611	0.8013
SEM	3.7512	3.7136	0.1547	0.0047	0.1135

NC = negative control, no additives in feed, PS = PoultryStar® me, AGP = antibiotic growth promoters, Bacitracin, ADG = average daily weight gain, FCR = feed conversation ratio, SEM = Standard Error of the mean, g = gram

**Table 10.** Effect of dietary treatments on growth performance of Ross 308 broiler chickens from day 1 to day 42, fed in the facility of Kasetsart University.

Treatment	Feed intake (g)	Body weight (g)	ADG (g/bird/day)	FCR	Livability (%)
NC	4740.160	2536.299	61.403	1.841	99.200
AGPs	4722.093	2572.408	62.196	1.811	99.733
PS	4724.067	2548.600	61.697	1.824	99.467
PS + AGPs	4751.960	2565.667	62.103	1.824	99.733
p -value	0.8165	0.3723	0.3732	0.2500	0.6368
SEM	12.4302	20.4605	0.4876	0.0115	0.1672

NC = negative control, no additives in feed, PS = PoultryStar® me, AGP = antibiotic growth promoters, Bacitracin, ADG = average daily weight gain, FCR = feed conversation ratio, SEM = Standard Error of the mean, g = gram

## CONCLUSION

Overall, the results of these two experiments under the controlled conditions proved that the evaluated synbiotic (PoultryStar® me) could serve as a replacement and an effective alternative to the Antibiotic Growth Promoters (AGPs), such as bacitracin and colistin in the broiler diets. With careful evaluation and the right preventive programs, the synbiotic can serve this purpose without being combined with AGP's. Hence, the replacement could be cost-effective and bring more value to broiler chicken producers.

## DECLARATIONS

### Authors' contribution

The experimental studies were conceived and designed by Basharat Syed and Yuwares Ruangapanit in consultation with Silvia Wein. Yuwares Ruangapanit supervised the experimental trials, collection of data, and its analysis. Silvia Wein reviewed the statistical analysis. The manuscript was written and drafted by Basharat Syed. All authors read and approved the final manuscript for submission and publication.

## Competing interests

The authors declare that they have no competing interests.

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## REFERENCES

- Applegate T, Klose V, Steiner T, Ganner A, and Schatzmayr G (2010). Probiotics and phytogenics for poultry: Myth or reality? *Journal of Applied Poultry Research*, 19: 194–210. DOI: <https://doi.org/10.3382/japr.2010-00168>
- Babazadeh D, Vahdatpour T, Nikpiran H, Jafargholipour MJ and Vahdatpour S (2011). Effects of probiotic, prebiotic and symbiotic intake on blood enzymes and performance of Japanese quails (*Coturnix japonica*). *Indian Journal of Animal Science*, 81: 870–874. Available at: <http://epubs.icar.org.in/ejournal/index.php/IJAnS/article/view/8799>
- Baldwin S, Hughes RJ, Van TTH, Moore RJ, and Stanley D (2018). At-hatch administration of probiotic to chickens can introduce

- beneficial changes in gut microbiota. *PLoS ONE*, 13(3): e0194825  
DOI: <https://doi.org/10.1371/journal.pone.0194825>
- Brennan J, Skinner J, Barnum DA, and Wilson J (2003). The efficacy of bacitracin methylene disalicylate when fed in combination with narasin in the management of necrotic enteritis in broiler chickens. *Poultry Science*, 82: 360-363. DOI: <https://doi.org/10.1093/ps/82.3.360>
- Brugaletta G, Cesare AD, Zampiga M, Laghi L, Oliver C, Zhu C, Manfreda G, Syed B, Valenzuela L, and Sirri F (2020). Effects of alternative administration programs of a synbiotic supplement on broiler performance, footpad dermatitis, caecal microbiota, and blood metabolites. *animals*, 10(3): 522. DOI: <https://doi.org/10.3390/ani10030522>
- Dhama K, Verma V, Sawant PM, Tiwari R, Vaid RK, and Chauhan RS (2011). Applications of probiotics in poultry: Enhancing immunity and beneficial effects on production performances and health: A review. *Journal of Immunology and Immunopathology*, 13: 1–19. Available at: [www.indianjournals.com/ijor.aspx?target=ijor:jii&volume=13&issue=1&article=001](http://www.indianjournals.com/ijor.aspx?target=ijor:jii&volume=13&issue=1&article=001)
- Food and Agriculture Organization (FAO) and World Health Organization (WHO) (2002). Working group report on drafting guidelines for the evaluation of probiotics in food. 30 April–1 May, London, UK, and Ontario, Canada. FAO, Rome, Italy. Available at: [https://www.who.int/foodsafety/fs\\_management/en/probiotic\\_guidelines.pdf](https://www.who.int/foodsafety/fs_management/en/probiotic_guidelines.pdf)
- Fuentes CG, Orozco LA, Vicente JL, Velasco X, Menconi A, Kuttappan VA, Kallapura G, Latorre J, Layton S, Hargis BM and Tellez G (2013). Effect of a lactic acid bacteria based probiotic Floramax-B11, on performance, bone qualities, and morphometric analysis of broiler chickens. An Economic Analysis. *Biological Systems*, 12(6): 322–327. Available at: <https://www.longdom.org/open-access/effect-of-a-lactic-acid-bacteria-based-probiotic-floramaxb11-on-performance-bone-qualities-and-morphometric-analysis-of-broilerchickens-an-economic-analysis-2329-6577-1000113.pdf>
- Gadde U, Kim WH, Oh, ST, and Lillehoj HS (2017). Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry: A review. *Animal Health Research Reviews*, 18: 26–45. DOI: <https://doi.org/10.1017/S1466252316000207>
- Gaggia F, Mattarelli P, and Biavati B (2010). Probiotics and prebiotics in animal feeding for safe food production. *International Journal of Food Microbiology*, 141: S15–S28. DOI: <https://doi.org/10.1016/j.ijfoodmicro.2010.02.031>
- Gustafson RH and Bowen RE (1997). Antibiotic use in animal agriculture A review. *Journal of Applied Microbiology*, 83: 531–541. DOI: <https://doi.org/10.1046/j.1365-2672.1997.00280.x>
- Huyghebaert G, Ducatelle R, and Van Immerseel F (2011). An update on alternatives to antimicrobial growth promoters for broilers. *The Veterinary Journal*, 187: 182–188. DOI: <https://doi.org/10.1016/j.tvjl.2010.03.003>
- Iji PA and Tivey DR (1998). Natural and synthetic oligosaccharides in broiler chicken diets. *World's Poultry Science Journal*, 54(02): 129–143. DOI: <https://doi.org/10.1079/WPS19980010>
- Koenen ME, Kramer J, van der Hulst R, Heres L, Jeurissen SH, and Boersma WJ (2004). Immunomodulation by probiotic lactobacilli in layer- and meat-type chickens. *Journal of British Poultry Science*, 45(3): 355–366. Available at: <https://www.tandfonline.com/doi/abs/10.1080/00071660410001730851>
- Kogut MH and Swaggerty CL (2012). Effects of prebiotics and probiotics on the host immune response. *Direct-Fed Microbials and Prebiotics for Animals*, Pages 61–72 Springer publications. Available at: <https://www.springer.com/gp/book/9781461413103>
- Kogut MH, Nan YX, Min YJ, and Broom L (2017). Gut health in poultry. CABI Wallingford UK, 12. Available at: <https://www.cabi.org/cabreviews/review/20173301742>
- Lee SI, Park SH, and Ricke SC (2016). Assessment of cecal microbiota, integron occurrence, fermentation responses, and Salmonella frequency in conventionally raised broilers fed a commercial yeast-based prebiotic compound. *Poultry Science*, 95: 144–153. DOI: <https://doi.org/10.3382/ps/pev322>
- Mead GC (2000). Prospects for 'competitive exclusion' treatment to control salmonellas and other foodborne pathogens in poultry. *The Veterinary Journal*, 159: 111–123. DOI: <https://doi.org/10.1053/tvjl.1999.0423>
- Mountzouris KC (2014). Probiotics as alternatives to antimicrobial growth promoters (AGPs) in broiler nutrition: modes of action and effects on performance. Probiotics in Poultry Production Concepts and Applications. Abdelrahman WHA, Mohnl M. editors, 5 m Publishing Ltd, Sheffield, UK, 129–157. Available at: <https://www.amazon.com/Probiotics-Poultry-Production-Concepts-Applications/dp/0955501180>
- Mountzouris KC, Palamidi I, Tsiroskos P, Mohnl M, Schatzmayr G, and Fegeros K (2015). Effect of dietary inclusion level of a multi-species probiotic on broiler performance and two biomarkers of their caecal ecology. *Animal Production Science*, 55: 484–493. Available at: <https://www.publish.csiro.au/an/an13358>
- Nava GM, Bielke LR, Callaway TR, and Castaneda MP (2005). Probiotic alternatives to reduce gastrointestinal infections: the poultry experience. *Animal Health Research Reviews*, 6(1): 105–118. DOI: <https://doi.org/10.1079/AHR2005103>
- Niewold TA (2007). The nonantibiotic anti-inflammatory effect of antimicrobial growth promoters, the real mode of action? A hypothesis. *Poultry Science*, 86: 605–609. DOI: <https://doi.org/10.1093/ps/86.4.605>
- Official methods of analysis (AOAC) (2016). 20th edition. Arlington: Association of official analytical chemists. Available at: [https://www.techstreet.com/standards/official-methods-of-analysis-of-aoac-international-20th-edition-2016?product\\_id=1937367](https://www.techstreet.com/standards/official-methods-of-analysis-of-aoac-international-20th-edition-2016?product_id=1937367)
- Oviedo-Rondon EO (2019). Holistic view of intestinal health in poultry. *Animal Feed Science and Technology*, 250: 1–8. DOI: <https://doi.org/10.1016/j.anifeedsci.2019.01.009>
- Pedroso AA, Menten JFM, Lambais MR, Racanicci A MC, Longo FA, and Sorbara JOB (2006). Intestinal bacterial community and growth performance of chickens fed diets containing antibiotics. *Poultry Science*, 85: 747–752. DOI: <https://doi.org/10.1093/ps/85.4.747>
- Pelicano ERL, Souza PA, and Souza HBA (2004). Productive traits of broiler chickens fed diets containing different growth promoters. *Brazilian Journal of Poultry Science*, 6(3): 177–182. Available at: <https://www.scielo.br/pdf/rbca/v6n3/a08v6n3.pdf>
- Roberfroid MB (1998). Prebiotics and synbiotics: concepts and nutritional properties. *British Journal of Nutrition*, 80 (Suppl. 2): 197–202. Available at: <https://pubmed.ncbi.nlm.nih.gov/9924284/>
- Saarela M, Mogensen G, Fonden R, Mättö J, and Mattila-Sandholm T (2000). Probiotic bacteria: safety, functional and technological properties. *Journal of Biotechnology*, 84(3): 197–215. DOI: [https://doi.org/10.1016/S0168-1656\(00\)00375-8](https://doi.org/10.1016/S0168-1656(00)00375-8)
- Snyder DL and Wostmann BS (1987). Growth rate of male germ-free Wistar rats fed ad libitum or restricted natural ingredient diet. *Laboratory Animal Science* 37: 320–325. Available at: <https://pubmed.ncbi.nlm.nih.gov/3613511/>
- Takahashi SE, MendesES AA, and Saldanha ESPB (2005). Efficiency of prebiotics and probiotics on the performance, yield, meat quality and presence of Salmonella spp in carcasses of free-range broiler chickens. *Brazilian Journal of Poultry Science*, 7(3): 151–157.

- Tayeri V, Seidavi A, Asadpour L, and Phillips CJC (2018). A comparison of the effects of antibiotics, probiotics, synbiotics and prebiotics on the performance and carcass characteristics of broilers. *Veterinary Research Communications*, 42: 195–207. Available at: <https://link.springer.com/article/10.1007/s11259-018-9724-2>
- Teirlynck E, Bjerrum L, Eeckhaut V, Huyghebaert G, Pasmans F, Haesebrouck F, Dewulf J, Ducatella R, and Immerseel FV (2009). The cereal type in feed influences gut wall morphology and intestinal immune cell infiltration in broiler chickens. *British Journal of Nutrition*, 102: 1453-1461. DOI: <https://doi.org/10.1017/S0007114509990407>
- Wise MG and Siragusa GR (2007). Quantitative analysis of the intestinal bacterial community in one- to three-week-old commercially reared broiler chickens fed conventional or antibiotic-free vegetable-based diets. *Journal of Applied Microbiology*, 102: 1138-1149. DOI: <https://doi.org/10.1111/j.1365-2672.2006.03153.x>
- Yang C, Cao G, Ferket P, Liu T, Zhou L, Zhang L, Xiao Y, and Chen A (2012). Effects of probiotic, *Clostridium butyricum*, on growth performance, immune function, and cecal microflora in broiler chickens. *Poultry Science*, 91: 2121–2129. DOI: <https://doi.org/10.3382/ps.2011-02131>
- Zhang Z and Kim I (2014). Effects of multi-strain probiotics on growth performance, apparent ileal nutrient digestibility, blood characteristics, cecal microbial shedding, and excreta odor contents in broilers. *Poultry Science*, 93: 364–370. DOI: <https://doi.org/10.3382/ps.2013-03314>