2023, Scienceline Publication *J. World Poult. Res.* 13(1): 1-19, March 25, 2023

Journal of World'^s Poultry Research

Review Paper, PII: S2322455X2300001-13 License: CC BY 4.0



DOI: https://dx.doi.org/10.36380/jwpr.2023.1

Poultry Management Strategies to Alleviate Heat Stress in Hot Climates: A Review

Shame Bhawa^(D), John Cassius Morêki^(D), and James Butti Machete^(D)

Department of Animal Sciences, Faculty of Animal and Veterinary Sciences Botswana University of Agriculture and Natural Resources, Private Bag 0027, Gaborone, Botswana

Corresponding author's E-mail: jmoreki@buan.ac.bw

Received: 08 January 2023 Accepted: 03 March 2023

ABSTRACT

Heat stress remains a major challenge affecting poultry production in sub-tropical and tropical environments; hence it continues to receive attention. The present study aimed to discuss heat stress and its effects on poultry production and suggests mitigation strategies to combat the effects of increased environmental temperature on poultry performance. Poultry raised in hot climates suffers from heat stress, which reduces meat and egg production, reproductive performance, feed intake, and feed conversion efficiency leading to poor growth rates. Reduced feed intake results in a reduction in meat quality, growth, egg yield, and quality. A decrease in feed utilization efficiency is the major cause of poor growth performance in hot environments. To counteract the negative impacts of high ambient temperatures on the performance of poultry, a wide range of management practices are widely used, including nutrient manipulations (particularly protein and energy), electrolyte and vitamin supplementation, feed form (especially particle size and moisture content), choice feeding, controlled feeding, time of feeding, wet feeding, water management, and use of new breeds that thrive well in hot environments. These management practices help lower heat load and facilitate evaporative cooling, all of which may positively impact poultry performance and health.

Keywords: Choice feeding, Feed conversion efficiency, Heat stress, Poultry production.

INTRODUCTION

There has been a significant increase in global average temperatures recently, which has affected the farming sector in the tropics (Barrett et al., 2019; Sohsuebngarm et al., 2019; Kennedy et al., 2022). High temperature above 32°C depresses feed intake, leading to poor performance in poultry (Cassuce et al., 2013; Sohsuebngarm et al., 2019). In addition, the increase in temperature results in the number of etiologically harmful microorganisms in the environment around the animals increasing. Due to an increase in parasites and microorganisms, climate change influences disease emergence and transmission (Ranjan et al., 2019). When the temperature in a living organism exceeds the threshold limit (i.e., thermo-neutral zone), it disrupts normal physiological functions and causes cell damage (Mack et al., 2013; Kennedy et al., 2022). High environmental temperatures typically cause stress-related issues such as output losses, metabolic alterations, poor development, and inefficiency (Dayyani and Bakhtiari, 2013; Afsal et al., 2018). At high temperatures, feed intake decreases while water intake increases (Mottet and Tempio, 2017).

Due to their insulating feathers, the absence of sweat glands on the skin, and the significantly high mass-to-body surface area ratio, broiler chicken strains are highly susceptible to rising temperatures (Scanes, 2015; Sejian et al., 2018; Bernabucci, 2019) compared to laying hens. In broiler chickens, for instance, rigorous genetic selection has enhanced metabolic activity in the pursuit of a higher development rate, further eroding the potential of a modern bird to withstand heat (Tamzil, 2014; Bohler et al., 2021). The broiler sector is confronted with heat stress, which raises production costs and degrades meat quality. This is attributable to the vulnerability of poultry to heat stress given the rapid metabolic and faster growth rates. In chickens, notably broilers, grown in hot climates, metabolic changes occur, resulting in a significant reduction in breast muscle growth (Safdar and Maghami, 2014).

Heat stress is divided into two types: acute heat stress (AHS), which is characterized by exposure to high temperatures for a short time, and chronic heat stress (CHS), which is characterized by exposure to high temperatures for a longer time (Lara and Rostagno, 2013; Pawar et al., 2016). In contrast to acute heat stress, chronic heat stress can increase fat content while destroying the muscles (Song and King, 2015; Adu-Asiamah et al., 2021). Besides the duration of excessive heat, the degree of heat stress influences the level of production (Adu-Asiamah et al., 2021). Both AHS and CHS have the potential to produce a significant decrease in poultry metabolism, which could lead to substantial issues with broiler growth performance and carcass characteristics which include meat color change, water holding capacity, muscular pH, and meat juiciness (Song and King, 2015; Gonzalez-Rivas et al., 2020).

Understanding the basic aspects underlying the causes, and impacts of heat stress, as well as, the approaches that can be used to mitigate or control such a widespread threat, will help solve worldwide food security challenges. Despite the ongoing debate in the literature on heat exposure, a synthesis of knowledge on such systems in terms of elevated ambient temperature exposure is still yet to be published. Therefore, this review aimed to discuss the management strategies that poultry producers can utilize to boost production in hot places around the world.

EFFECTS OF HEAT STRESS ON POULTRY

Heat stress (also referred to as hyperthermia) is a result of global warming and is considered one of the crucial factors that negatively influence poultry production (Vandana and Sejian, 2018). Excessive heat depresses feed intake, feed conversion efficiency, growth, meat and egg output, and reproductive function (Alverdy and Luo, 2017; Quinteiro-Filho et al., 2017; Rostagno, 2020). The reduced feed intake due to high temperatures has a negative effect on semen quality and fertility, thus leading to poor hatchability rates (Nawab et al., 2018; Nyoni et al., 2019). In addition, heat stress affects a poultry's production performance. digestive health, body temperature, immunological responses, hunger hormone modulation, and oxidative properties (Goel, 2021). Recently, Nawaz et al. (2021) observed that heat stress degrades meat quality by altering the pH, water-holding capacity, and drip loss in the meat leading to changes in the normal meat color, flavor, and texture of chicken meat. Moreover, the effects of heat stress on meat quality include a reduction in protein synthesis and an increase in unfavorable fat (Kadykalo et al., 2018). By adjusting to changing climatic conditions, poultry frequently sacrifices their production capacity (Slawinska et al., 2019; Smith et al., 2019). However, poultry breeds are more resilient to climate change which continues to influence egg and meat production (Farag and Alagawany, 2018; Liverpool-Tasie et al., 2019).

Overcrowding and high outside temperatures contribute to the development of heat stress. However, by increasing cooling options, which include using the fogging system, use of a wet pad system, and microsprinklers, the heat load may be reduced by lowering the heat production level or changing the pattern of thermal production throughout the day (Gicheha, 2021). Commercial broilers' growth rate and meat yield are known to be slowed by high ambient temperatures (Zhang et al., 2017). In addition to poorer weight gain, high mortality rate, and reduced feed consumption, high temperatures negatively affect intestinal development (Rostagno, 2020). Furthermore, high temperatures disrupt broilers' acid-base balance and increase respiratory rate which can contribute to respiratory alkalosis (Scanes, 2015).

Heat stress can have a substantial influence on layer flocks, but some precautions can be done to keep hens healthy and produce eggs. For instance, the lighting schedule should be changed to provide more light hours during the colder hours of the day to promote feed consumption during cooler times of the day. In addition, when it is hot outside, it is best to lower stocking density (Reddy and Ramya, 2015; Abbas et al., 2021). High stocking rates during the hot season can lead to inadequate ventilation. Early heat conditioning also appears to be an effective method for boosting the heat tolerance of some chicken breeds (Saeed et al., 2019). Layer flocks can be kept calm by starving or fasting during hot hours (Saeed et al., 2019; Bilal et al., 2021; Shakeri and Le, 2022). Therefore, egg producers must be prepared when summer temperatures rise as egg yield will decrease and flock mortality increases (Yahav, 2015; Sinha et al., 2018).

During the chicks' first few days of life, chickens cannot regulate its heat production in response to the environmental temperature, therefore a decrease in environmental temperature leads to a reduction in body temperature (Ranjan et al., 2019). However, after 21 days, chicks start to develop additional homeothermic traits, such as the capacity to match their heat production to the surrounding temperature, allowing them to endure the lowering effect that a decrease in ambient temperature has on their body temperature (Ranjan et al., 2019; Saeed et al., 2019). The normal body temperature of an adult chicken is 40.6-41.7°C (Ranjan et al., 2019). The comfortable ambient temperature for adult poultry is 18-24°C, whereas chicks require higher temperatures of around 32°C in their first week of life which decreases over time (Scanes, 2015). Above 32°C, poultry fails to maintain their normal internal body temperature, due to the absence of sweat glands and the presence of complete feather coverage of the body (Hu et al., 2016). When the ambient temperature rises above 24°C, the internal body temperature of the chicken also rises, which causes it to consume less feed (Cassuce et al., 2013). Heat stress, panting, and prostration results at a temperature above 35° C (Hu et al., 2016). When a chicken's core body temperature reaches a critical level of 47°C, sometimes known as the upper lethal temperature, chickens may die from heat prostration (Reddy and Ramya, 2015; Scanes, 2015). In laying hens, heat stress causes low egg production and an increased number of hatching egg rejects in breeder hens (Abbas et al., 2021). Heat stress is less likely similar to affect younger and lighter chicks than older and heavier chickens (Farag and Alagawany, 2018). Therefore, heat stress can be alleviated by modifying the macro and microenvironments in which chickens are kept. High humidity and high environmental temperatures adversely affect poultry production (Saeed et al., 2019; Yousaf et al., 2019).

POULTRY RESPONSES TO HEAT STRESS

The susceptibility of broiler chickens to heat stress increases as air relative humidity and ambient temperature

values are above the thermal comfort zone (16-23°C and 50-70% relative humidity), making it hard for birds to release heat (Gamba et al., 2015). This results in their body temperature rising, which harms their growth performance. Hot weather causes poultry to perform poorly as it results in decreased feed intake and increased water intake (Saeed et al., 2019; Rahman and Hidayat, 2020). At high temperatures, laying hens lay fewer eggs, watery eggs, and eggs with thin shells or even shell-less eggs due to lack of calcium; grow slower; and are more likely to become sick due to their compromised nutritional requirement as protein digestibility is reduced up to 9.7% (Habashy et al., 2017; Nawaz et al., 2021). In broiler chickens, decreases in growth rates, feed efficiency, immunity, and carcass quality were observed at high ambient temperatures (Dayyani and Bakhtiyari, 2013). Aswathi et al. (2019) reported a reduction in fertility percentage (-7.22%) and hatchability of fertile egg sets (-2.51%) in breeders. Heat stress has a negative effect on not just feed intake and utilization, but also carcass quality (Rath et al., 2015, Aswathi et al., 2019; Rahman and Hidavat, 2020) due to the unfavorable partitioning of metabolizable energy consumed, with a large proportion of it being stored as fat and the remainder as muscle (Rahman and Hidayat, 2020). The signs of a heat-stressed chicken include panting, extending the wings, holding the wings slightly apart from the body, standing or lying down, and closing the eyes (Dayyani and Bakhtiyari, 2013). A study by Altan et al. (2003) reported that heat stress increases fearfulness, induces oxidative stress, and initiates significant physiological responses in broiler chickens. Birds can survive a gradual increase in temperature, but a rapid increase in temperature will result in higher mortality rates (Rostagno, 2020). Figure 1 illustrates the responses of chickens to heat stress.



Figure 1. Poultry responses to heat stress

BIOLOGICAL CHANGES IN CHICKENS DUE TO HEAT STRESS

Heat stress in poultry results in several behavioral, physiological, and neuroendocrine changes that influence health and performance (Ahmad et al., 2022). The major physiological changes that occur in heat-stressed poultry are discussed briefly below.

Oxidative stress

Reactive oxygen species (ROS) are peroxyl radicals produced by cells during normal metabolism and are required for physiological functions such as ion transport, immunomodulation, and cytokine production (Wasti et al., 2020). Extra ROS is removed from cells through physiological detoxification processes. The Nrf2, a transcriptional factor when activated, under thermoneutral conditions, causes an increase in the production of a collection of antioxidant molecules that deal with the elevated ROS generated within the cell (Surai et al., 2019). Since the mitochondria create a significant amount of ROS, excessive mitochondrial ROS production may be a key factor in oxidative stress. Acute heat stress increases the formation of ROS from mitochondria, harming birds' skeletal muscles by oxidation (Akbarian et al., 2016). Acute heat stress causes an increase in the activity of the electron transport chain and mitochondrial substrate oxidation, which results in an excessive generation of superoxide (Akbarian et al., 2016).

Heat stress has been linked to cellular oxidative stress in chickens (Estévez, 2015; Surai et al., 2019). Down-regulation of chicken uncoupling protein exacerbates the oxidative stress situation during the later stages of acute heat stress, leading to mitochondrial malfunction and tissue damage (Mishra and Jha, 2019). Constant heat stress reduces the mitochondria's ability to create oxidative energy and consequently increases the chicken's uncoupling protein, this significantly alters the pattern of antioxidant enzyme activities, leading to the depletion of antioxidant reserves (Sahin et al., 2016). Oxidative stress has been linked to reduced growth rates, biological defects, loss of income, and severe health concerns in poultry (Estévez, 2015; Zaboli et al., 2019). While the chicken's physiology struggles to maintain thermal homeostasis, elevated ROS concentrations increase in stressful environmental situations (Sahin et al., 2016). In an effort to defend itself from the damaging effects of ROS on cells, the body undergoes an oxidative stress state and starts to manufacture and release heat shock proteins (HSP, Archana et al., 2017). Studies by Arnal and Lallès (2016) and Hao et al. (2017) have demonstrated that when exposed to heat stress, laying hens and broilers have higher *HSP70* levels.

Role of genes in heat stress

The global poultry industry has difficulty with genetic screening for high-temperature tolerant broilers (Zeferino et al. 2016). Therefore, crossing commercial chickens with strains that are highly tolerant to high temperatures can also be used to integrate heat stress resistance into the genome. The most common breeding approach for generating a commercial hybrid robust to tropical conditions and capable of producing a respectable amount of eggs and meat is a crossbreeding program between indigenous and foreign breeds (Duangjinda et al., 2017; Abd El-Hack et al., 2018).

The introduction of genetics from high-temperature tolerant strains into grandparental stock is a useful technique for speeding up the genetic advancement of commercial strains that can withstand heat stress. In chickens, heat-tolerant genes such as dwarfism (Vandana et al., 2021), naked-neck (Desta, 2021), slow/rapid feathering (Wells et al., 2012), and frizzle genes (Fathi et al., 2013; 2019), have been extensively studied. In every case, the chickens' appearance and performance indicators which include body weight gain, body weight, and feed conversion ratio (FCR), were influenced by their genes (Nawaz et al., 2021). Another downside of temperature control is immune inhibition (Goel et al., 2021). When exposed to high ambient temperature, differences in the levels of several immunological marker genes including interleukins (ILs), tumor necrosis factors (TNF), and tolllike receptors (TLRs) had a more pronounced increase in the spleen and intestine of chicks (Varasteh et al., 2015; Moraes et al., 2019).

The response of prokaryotic and eukaryotic cells to potentially harmful stimulations like heat stress induces the synthesis of stress proteins which are referred to as heat shock proteins (Efeoğlu, 2009). Many strategies, including the development of thermotolerance, modification of apoptotic and anti-apoptotic signaling pathways, and control of cellular redox conditions, are used by heat-shock proteins to provide protection against heat stress to cells (Shehaha et al., 2020). The HSP70 and HSP90 relate to families of HSP that are around 70 and 90 kilo Daltons, respectively (Datta et al., 2017). The HSP70 gene is thought to protect the body from the harmful consequences of oxidative stress (Xie et al., 2015), whereas *HSP90* engages with client proteins during the last stages of folding and changes their shape (Kumbhar et al., 2018). The *HSP70* is a chaperone polypeptide that successfully protects a variety of proteins and cell components from stress (Habashy et al., 2017; Perin et al., 2021). In chickens, Cedraz et al. (2017) found a nucleotide polymorphism in the coding area of *HSP70*. Hyperthermia causes oxidative stress and promotes the formation of ROS, resulting in the stimulation of *HSP70* expression (Robert et al., 2017).

Acid-base balance

As the ambient temperature rises, birds must release heat through panting as thermoregulation is difficult (Wasti et al., 2020). Panting is a behavior in which chickens open their beaks to increase their rate of breathing such that the respiratory tract will provide the evaporative cooling effect (Park and Kim, 2016). When panting occurs, CO₂ is excreted faster than it is produced by the cells, causing the blood's regular bicarbonate buffer system to be disrupted. Carbonic acids (H₂CO₃) and hydrogen ions (H^+) concentrations decrease when CO₂ levels are reduced (Hamm et al., 2015). On the other hand, the concentration of H₂CO₃ is raised leading to an increase in the blood pH, and the blood becomes alkaline. To cope with this situation and maintain a normal blood pH, chickens will begin to expel more H₂CO₃ and retain H⁺ from the kidneys (Saeed et al., 2019). The increased H⁺ disrupts the acid-base balance, resulting in respiratory alkalosis and metabolic acidosis, as well as, a reduction in poultry production (Zaboli et al., 2017).

Suppressed immune-competence

Chickens pant to expel heat and reduce body temperature, but they frequently experience instabilities in their overall energy balance because of insufficient feed consumption under heat stress (Hirakawa et al., 2020). In broilers, the weights of the main organs such as the liver and pectoral muscle do not improve as expected under the heat stress situation, in addition to impairment of broiler growth performance (Piestun et al., 2017; Hirakawa et al., 2020; Tang et al., 2022). Decreased humoral immunity is one of the most common forms of immunodeficiency in heat-stressed chickens, which might increase the risks of secondary infections that restrict vaccination efficacy (Lara and Rostagno, 2013).

Bursa of Fabricius is a fundamental immunological tissue unique to birds that are connected to the cloaca and it is necessary for B cell development and antibody competence diversification brought on by gene conversion and V(D)J recombination that causes B cell exportation to the lower limbs (Ratcliffe et al., 2014; Monson et al., 2018). Continuing heat stress accelerates the rate of the bursa of Fabricius atrophy and adds to the atrophy of the other immune components in hens intensively selected for muscle yield and growth (Jahanian and Rasouli, 2015; Campbell et al., 2019). Reduced intestinal integrity, which increases exposure to pathogens and antigenic compounds such as lipopolysaccharides (LPS), or systemic stress responses such as circulatory cytokines and acute-phase proteins, could affect the bursa of Fabricius during heat stress (Nochi et al., 2018). These factors influence the formation, survival, and motility of the bursa of Fabricius (Calefi et al., 2016).

The hypothalamic-pituitary-adrenal and sympathetic adrenal medullar axis are the main mechanisms by which the body's immune response can be altered (Herman et al., 2016; Goel et al., 2021). Neuroendocrine products of both hypothalamic-pituitary-adrenal the and sympathetic adrenal medulla axes including cortisol and catecholamines have been shown to have receptors on monocytes, lymphocytes, and granulocytes, which might affect proliferation, cytokine production, cellular trafficking, cytolytic activity, and antibody production (Bohler et al., 2021). Heat stress affects the microbiome's makeup and abundance in addition to causing oxidative stress in the gut epithelium, which impairs permeability and increases susceptibility to infection and inflammation (Cao et al., 2021).

It has been shown that broilers that have been exposed to heat stress had decreased concentrations of free circulating antibodies and specific IgG and IgM, along with lower levels of general and humoral reactivity (Van Goor et al., 2017). The weights of the bursa, thymus, liver, and spleen were also observed to be drastically lowered. Similarly, Cantet et al. (2021) reported reduced bursa weight and lymphocyte numbers in the medulla and the cortex regions of the bursa in broilers exposed to heat stress. Faud et al. (2016) also reported that heat stress was associated with a decrease in spleen and thymus size in laying chickens. Heat stress has also been shown in recent research to change the number of circulating cells (Santos et al., 2015). Due to lower quantities of circulatory lymphocytes and greater concentrations of heterophils, heat stress leads to a significant increase in the heterophil: lymphocyte ratio which is an indication of chronic stress (Santos et al., 2015; McGregor et al., 2016). Consequent to this, communicable and infectious poultry diseases such as Newcastle and infectious bursal disease become more prevalent in tropical environments throughout the summer

(Badruzzaman et al., 2015; Saelao et al., 2021). In another study, Hirakawa et al. (2020) reported lowered levels of antibodies in heat-stressed birds (Hirakawa et al., 2020).

Neuroendocrine changes

During heat stress, the neuroendocrine system is critical for the maintenance of homeostasis and proper physiological functioning in poultry (Jessop et al., 2016). The sympathetic nerves detect a rise in ambient temperature and send an impulse to the adrenal medulla (Kumari and Nath, 2018), which enhances catecholamine secretion in response to stress (Ruuskanen et al., 2021), resulting in elevated blood glucose levels, exhaustion of liver glycogen, loss of muscle glycogen, accelerated respiration rate, peripheral blood vessel vasodilation, and heightened neurological responsiveness (Kumari and Nath, 2018; Beckford et al., 2020). In response to stress, the hypothalamus releases a corticotrophin-releasing hormone (CRH), which induces the pituitary to release adrenocorticotrophic hormone (ACTH, Wasti et al., 2020). Corticosteroids are produced and released by the adrenal glands in response to ACTH (Souza et al., 2016). Corticosteroids increase plasma glucose levels by stimulating gluconeogenesis (Kumari and Nath, 2018). The thyroid thyroxine and hormones triiodothyronine are also crucial in maintaining a consistent metabolic rate by playing a pivotal role in digestion, and heart and muscle function (Cioff et al., 2013; Wasti et al., 2020).

HEAT STRESS MITIGATION STRATEGIES

The strategies to combat heat stress are categorized broadly as genetic approach, managerial practices, and nutritional manipulation.

A genetic approach to mitigate heat stress

Current developments in genetics and biotechnology may pave the way for the investigation of changes to the chicken gene to assist reduce heat stress (Cedraz et al., 2017). The increased metabolic rate of improved broiler lines makes them more sensitive to heat stress. Therefore, improving the production qualities of these breeds in hot and arid environments may require creating poultry lines that incorporate some of the genes that reduce heat stress (Wasti et al., 2020).

A single dominant autosomal gene called "naked neck" enables chicken necks to have less plumage, which helps the neck to dissipate heat (Tóth et al., 2021). In heterozygous necked neck (Na/na) and homozygous necked neck (Na/Na), the naked neck gene reduces the neck plumage cover by 20% and 40%, respectively, in comparison to normal siblings (*na/na*, Rajkumar et al., 2010). In broilers, the *Na* gene is associated with an increase in body weight and breast muscle, a decrease in abdominal fat, and an increase in body temperature (Wang et al., 2018). It was found that the heterophil to lymphocyte (H/L) ratio and total plasma cholesterol levels of the naked-necked chickens were much lower throughout the hot season compared to normal chickens (Wasti et al., 2020). Under high temperatures, laying hens with the bare neck gene also demonstrated improvements in egg weight, number, and quality (Azhar et al., 2019). These experiments show that it is possible to use these genes to create a breed of chicken that can withstand heat stress.

The *frizzle* (F) gene enables the feather's edge to curve, which decreases the feather's weight, enhances heat radiation from the body, and improves the feather's ability to act as an insulator (Nawaz et al., 2021). Relative to heterozygous carriers and regular feathered hens, laying hens with the homozygous frizzle gene had increased egg production and quality features by enhancing the extent of heat dissipation (Kumari and Nath, 2018). Except for sexual development under heat stress, there is a positive interaction between the feathering genotype (FF) and ambient temperature for all reproductive variables, including egg production, hatchability, and chick production (Dong et al., 2018).

In poultry, a sex-linked recessive gene called the dwarf gene (dw) causes homozygous females and males to weigh about 30% and 40% less than normal, respectively (Zerjal et al., 2013). The benefit of the dw gene in heat-stressed laying hens has been the subject of some debate (Wasti et al., 2020). However, Fathi et al. (2022) recommended the development and commercialization of *frizzled* and *naked-necked*, and *dwarf* genes in poultry.

Managerial practices *Housing*

In the tropics, poultry houses are predominantly naturally ventilated open-sided (Alchalabi, 2013). With rising air velocity, heat loss via radiation and convection can increase significantly (Saleeva et al., 2019; Elbaz et al., 2021). Therefore, it is best to allow natural airflow from the north and south sides while also shielding birds from direct sunlight throughout the day; thus, the shed's longitudinal direction should be from east to west (Oloyo and Ojerinde, 2019). To maintain their internal temperature, poultry houses should be designed with optimal insulation (Scanes, 2015).

The roof of the poultry shed should be at a 45° angle which will be able to maximize the distance of the poultry from the heat accumulated under the roof (Olovo and Ojerinde, 2019). Furthermore, water sprinkling can keep the roof cool at high temperatures (Saeed et al., 2019). The heat that is gained or lost from the building is significantly influenced by the size, pitch, the roof's color, reflectivity, and direction, as well as, the structure's ventilation system (Wang et al., 2018). According to Saleeva et al. (2019), the reflectivity of the roof can be increased by adding an aluminum roof or painting it with metallic zinc. Evaporative cooling technologies with cooling pads and sprinklers inside the chicken house can be used in farms with extreme outside temperatures and low relative humidity (Saeed et al., 2019). Glass wool is currently used as an insulating material in the ceiling of environmentally controlled chicken houses (Alchalabi, 2013).

Stocking density is another factor that contributes to heat stress. A study by Moreki et al. (2020) in Botswana showed that the stocking density of 10-12 $birds/m^2$ was ideal for open-sided poultry sheds in summer. The authors concluded that broiler chicken growth performance was negatively impacted by stock densities of more than 12 birds/m². In another study, Gholami et al. (2020) reared broilers at four different stocking densities (10, 15, 17, and 20 birds/m²) under hot and dry conditions and observed that the stocking density of 10 birds/m² resulted in lower FCR, higher body weights, weight gains, and feed intake compared to those reared at 15, 17 and 20 birds/m². The higher metabolic rate of chickens during the summer increases heat generation inside the poultry house and slows heat loss during hot and humid weather giving rise to an increase in the poultry house's total temperature (Nilsson et al., 2016; Donald, 2018).

The use of corrugated iron sheets and walls which are painted white to reflect heat is encouraged in subtropical and tropical regions (Olovo and Ojerinde, 2019). Furthermore, grass can be used as a roofing material which can also serve as an insulation material. Sidewalls should have roll-down reinforced curtains that can be adjusted for use in cold weather and at night (Bhadauria, 2017). A sidewall's height should be between 25 and 70 cm high to allow natural airflow during the hot period as side wall curtains will be rolled down (Oloyo and Ojerinde, 2019). The open space between the sidewall and the roof gable will be closed with a 25 mm wire mesh (Alchalabi, 2013). However, as technology progresses, the use of a closed housing system for the intensification of agricultural operations has increased significantly (Donald, 2018). Climate-controlled housing systems (also referred to as closed buildings) with exhaust fans, air conditioning, cooling pads, and cool perches are beneficial in assisting chickens in dealing with the negative consequences of heat stress (Bhadauria, 2017). Closed buildings, on the other hand, are costly to construct and maintain (Glatz and Pym, 2013).

Feeding strategies

Only feeding methods can lessen heat exhaustion if the animal generates less heat or dissipates heat from the body through radiation during tunnel ventilation, where air velocity is higher. Lower heat production can be realized by a reduced heat increment, catabolism of fewer nutrients above requirements, or more efficient nutrient digestion (Barrett et al., 2019). Broiler chickens compared to laying hens appear to need more attention to feeding schedules. Many of the difficulties related to heat exhaustion in broilers can be alleviated simply by feeding at the right time (Syafwan et al., 2011; Kennedy et al., 2022). To address heat stress, coarser meals, diurnal feeding patterns, self-selection procedures, and wet feeding are all viable options. The feed should be well processed into mash, crumb, or pellets, and supplementary feeders should be available on hot days to increase appetite (Rahman and Hidayat, 2020).

The use of low-beam lights may also minimize activity, thus lessening the heat burden on the birds (Bhadauria et al., 2016). Lighting schedules are utilized for broiler chickens to control feed intake (Wu et al., 2022) and provide access to feed and water, especially during the cooler parts of the day (De Oliveira and Lara, 2016). The length of the photoperiod can be altered as an alternate strategy to enhance the well-being, immune response (Riber, 2015), and ultimately the performance of birds that are under heat stress (Parvin et al., 2014). Using low-intensity lighting when the temperature is high (for example 180 Lux) can prevent broilers from moving around and agitating, which can lead to them to be heavier (Mousa-Balabel et al., 2021; Wu et al., 2022). Mousa-Babel et al. (2021) compared the performance of broiler chickens reared at low-beam blue light intensity (5 Lux), medium blue light (20 Lux), and high blue light intensity (320 Lux) and found that broiler chickens raised under low-beam blue light intensity had significantly higher body weight, body weight gain, antibody titers against the Newcastle disease virus, and foot pad dermatitis compared to their counterparts in high blue light intensity. In addition, chickens on low-beam blue light intensity had lower activity levels and heterophil/lymphocyte ratios, and FCR.

Feeding time is a significant component in reducing the effects of heat stress on feed intake and utilization (Farghly et al., 2018, Kennedy et al., 2022). Therefore, during the time of low temperatures, for example, in the early hours of the day and late evening, a significant portion of the feed should be supplied to the poultry, with the remaining amount available *ad libitum*. According to Daghir (2009), chickens that are feed-starved produce less heat than those that are fed; hence removing feed on hot days has some ameliorating benefits on performance. Farghly et al. (2018) reported that feed withdrawal involves alterations in intestinal morphology and depletion of intestinal mucosa due to fasting which may damage the intestinal cells.

A study by Zaboli et al. (2019) reported that a rise in the room temperature from 21.1°C to 32.2°C leads to a decline in feed intake of around 9.5% /bird/day from 1 to 6 weeks of age. In another study, He et al. (2019) reported that a rise in environmental temperature from 32.2°C to 37.8°C results in a 9.9% decrease in feed intake per bird/day. It is, however, not recommended to allow birds to go for a long time without a feed as this will have an impact on growth and may increase skin scratches at feeding time resulting in downgraded carcasses (Suganya et al., 2015; Vandana et al., 2021).

The form in which the feed is presented to the birds affects the consumption of poultry exposed to high environmental temperatures. In warmer conditions, poultry, particularly broilers, prefer eating larger particles (Ranjan et al., 2019; Massuquetto et al., 2020). According to Smalling et al. (2019), when broilers are fed pelleted feed, the energy required for feeding is reduced by 67%, allowing that energy to be channeled toward more productive applications. Khalil et al. (2021) reported that feeding pellets to laying hens during high ambient temperatures contributes to higher feed efficiency, egg production, and water intake compared to mash feeding. The physical feature of the pellets enables the birds to ingest feed with less wasted energy, therefore the pellets' quality and durability are extremely important. The FCR can be altered by 0.01 points if the pelleted feed contains 10% fine particles (Ahmed and Abbas, 2013).

The quantity of coarse particles in droppings is adversely correlated to the water in the droppings. The higher retention duration of coarse particles inside the gastrointestinal tract (GIT) is responsible for this association (Smalling et al., 2019; Abdel-Moneim et al., 2021). In comparison to fine diets, coarse diets can enable more retention of water from GIT (Smalling et al., 2019) and this may aid to release the heat burden. More heat loss by evaporative cooling, on the other hand, emphasizes the importance of increased water intake in heat-stressed birds (Lara and Rostagno, 2013). Therefore, the provision of high-physical-quality feed will minimize energy expended and heat generated during feeding (Mir et al., 2018).

Choice feeding encourages chickens to select a meal and reduce the heat burden associated with the metabolic process in hot environments. It could also help the chickens to better match their nutrient intake to their needs. When given a choice of diet, chickens are reported to select a variety of food items to meet their nutrient needs (Sinha et al., 2018). It has been observed that chickens that are choice fed choose feed ingredients with lower heat increments to minimize excess heat during the harshest times of the day, thus enhancing their heat tolerance (Diarra et al., 2014). Suganya et al. (2015) reported that choice-fed broilers ingest less protein and much more energy at high temperatures than those feeding on a complete diet, presumably to limit body heat output from protein-high heat increment. Similarly, De Almeida et al. (2012) observed that when Japanese quails were kept at temperatures ranging from 20 to 35°C, they chose to eat more calories and less protein when given a choice diet vs. a single complete diet.

Diet management changes, including rehydrating feed, have long been known to improve poultry performance (Rahman and Hidayat, 2020). Relative to broiler chickens consuming dry feed, this technique enhances weight gain, feed intake, FCR, and the weight of the gut in broilers at ordinary temperatures (Kaldhusdal et al., 2016; Rostagno., 2020). In another study, it was reported that even though the weight of the digesta across the entire digestive system of chickens was lower while the feed intake was higher, wet feeding has been associated with a quicker rate of passage through the gut (Calefi et al., 2016).

A previous study by Calefi et al. (2014) reported advancements in digestive efficiency which were assumed to be due to a higher empty weight, a longer gut length, and greater gut wall thickness in some areas of the digestive tract with wet feeding. Farghly et al. (2018) and Kadykalo et al. (2018) observed that wet feeding increased the ingesta's fluidity, possibly indicating a faster digesta transit rate. Additionally, a thicker intestinal wall could help with digestion. Farghly et al. (2018), compared rehydrating to dry feed and found that rehydrating feed lowers digesta fluidity to a similar degree and promotes pre-digestion and absorption, presumably due to faster digestion enzyme penetration into feed particles. This may result in increased nutrient digestibility. In addition, external enzyme inclusion in the wet feed may have an additional potential influence on absorption, since they may promote substrate accessibility for enzymes, hence, increasing nutrient absorption (Holtmann et al., 2017). Saleh et al. (2021) reported that wet feeding may improve performance because it increases dry matter (DM) intake at high temperatures. Egg weight and egg production could be boosted in this manner under high temperatures. Waiz et al. (2016) observed that compared to dry feeding, moistening laying hen's feed at a 1:1 (feed: water) ratio in hot environments improves laying performance. High performance in hot conditions is predominantly caused by an increase in DM intake on wet feed, which enhances the intake of micronutrients (Afsharmanesh et al., 2016).

At high temperatures birds eat less, thus failing to meet their nutrient needs (Rath et al., 2015). Therefore, heat stress can be alleviated by increasing the nutrient density of the diet. During summer, especially for broiler hens, adding fat to the diet should be taken into consideration to keep their daily energy requirement in line with their needs for growth (Diarra and Tabuaciri, 2014; Teyssier et al., 2022). Due to fat's lower heat increment when compared to alternative energy sources like carbohydrates or proteins, the inclusion of fat in diets for broilers that are under heat stress improves their feed intake and performance (Rath et al., 2015; Pursey et al., 2017). However, to achieve a balanced meal and hence optimize utilization, the content of other nutrients, notably proteins, must be appropriately adjusted whenever the energy density is raised by added fat (Rahman and Hidayat, 2020). Heat-stressed chickens have a strong urge to reduce feed intake to lower their body temperature (Wasti et al., 2020). Low-digestible energy and proteinrich diets are favorable when heat stress is moderate (Lemme et al., 2019). In addition, it was reported that fat in the diet increases nutrient utilization by slowing feed passage through the GIT (Jha and Mishra, 2021).

According to a previous study, polyunsaturated fatty acid-rich fat sources, such as soybean oil, fish oil, canola oil, flaxseed oil, and walnuts must be avoided or be used in moderation, indicating that caution must be exercised when choosing a fat source to include in a diet (Seifi et al., 2020). According to Surai et al. (2019), such sources are deficient in antioxidants and are vulnerable to oxidative rancidity, which results in the degradation of vitamins A and E and the taste of poultry meat being altered. Moreover, soybean oil has a high concentration of polyunsaturated fatty acids that frequently result in the creation of excess visceral and breast intramuscular fat, which lowers the quality of the carcass (Abdel-Moneim et al., 2021). However, if the energy density of the diet is to be increased, the levels of all nutrients must be adjusted to maintain optimal intake (Pawar et al., 2016).

It was previously noted, poultry limit feed consumption in hot weather which results in nutrient deficiencies (Teyssier et al., 2022). Due to a reduction in consumption, there is a decrease in the intake of essential nutrients, such as protein, essential amino acids, minerals, and vitamins (Rath et al., 2015). In this case, it is preferable to improve and balance vital amino acids because increasing protein levels can increase heat production during protein metabolism (Teyssier et al., 2022). Bird performance is unaffected even if the diet is lacking in protein but contains a balanced amino acid content (Kumar et al., 2016). If protein levels must be increased, vegetable-derived proteins such as soy, sesame, and sunflower are excellent choices since animal-source proteins will produce more heat during metabolism (Tari et al., 2020). Vegetable proteins are rich in arginine, an essential amino acid required during heat stress. Dao et al. (2021) reported that the role of arginine aids in protein synthesis and immunity. At the macrophage level, arginine is transformed into nitric oxide (Rath et al., 2014), a mediating component in vasodilation and increased peripheral blood flow which are significant thermoregulatory responses to heat stress (Liu et al., 2019).

When feed intake is lowered due to heat stress, it was normally advised that dietary protein levels be raised to maintain a steady protein intake (Liu et al., 2019). However, studies conducted over time suggest that birds under heat stress may not always require more protein (Suganya et al., 2015). A recent study reported that feeding broilers high-protein diets at high environmental temperatures result in their growth being inhibited (Qaid and AlGaradi, 2021). It was indicated that hens' growth performance at 3 to 6 weeks of age under hot temperatures of 32°C was not improved by raising protein content from 17 to 23% (Awad et al., 2019). This was primarily caused by the increased nitrogen excretion and reduced efficacy of the high-protein diet compared to the low-protein diet (Kidd et al., 2021). Previous investigations demonstrated that higher body heat generation due to increased feed intake contributed to poor performance (Diarra et al., 2014). The mentioned authors reported that birds on lowprotein diets consumed more protein, possibly due to a physiological shift that allows them to use the protein more efficiently when it is scarce. On the other hand, prolonged heat-stress exposure affects the reaction of poultry to dietary protein levels, therefore lowering crude

protein levels as a strategy for mitigating heat stress is not justified (Bohler et al., 2021).

In a separate study, it was found that using protein sources that provide the appropriate amounts and proportions of methionine and lysine can lower 2-4% of dietary protein without compromising weight increase or feed conversion (Attia et al., 2020). It was reported that adding 0.05% methionine to water boosted feed efficiency considerably in heat-stressed chickens (Cadirci and Koncagul, 2014). Any loss in amino acids will result in their insufficiency, making protein non-ideal irrespective of the protein amount (Kumar et al., 2016). Therefore, supplementing low-protein meals with essential amino acids has been shown to help heat-stressed chickens perform better (Lemme et al., 2019). Heat intensity and duration, breed, age of birds, the quantity of amino acid supplementation, and diet composition could all influence how heat-stressed chicken responds to low-protein diets. Under hyper thermoneutral conditions compared to thermoneutral conditions, the total sulfur amino acids (TSAA) demand would be higher (Babazadeh and Ahmadi, 2022). In addition, it takes more TSAA to attain optimal growth performance when broiler chicks are raised at high temperatures (Del Vesco et al., 2013; Zarghi et al., 2020). When adding methionine supplements, factors such as age and production parameters must be considered to mitigate the harmful effects of heat stress (de Freitas Dionizio et al., 2021). Supplementing with methionine is also useful for lowering immunological stress and can change how the immune system responds (Pacheco et al., 2018).

Feed as a source of calcium carbonate

Calcium is supplied to commercial breeders in many ways which include using grower diets that contain 0.9 to 1.0% calcium supplemented with up to 5% egg production or using classical pre-breeder diets that allow for the development of greater medullary bone reservoirs without using the diets that contain 2-2.5% calcium (Bryden et al., 2021). Heat stress causes poultry to consume less than 3.5 grams of calcium each day (Abbas et al., 2021). In addition, heat stress decreases the production of calbindin, a calcium-binding protein required for calcium absorption in the intestine (Ebeid et al., 2012). Ranjan et al. (2019) it is reported that feeding laying hens in the evening improves their laying rate and eggshell quality by increasing calcium intake. A decrease in egg production is directly linked to a reduction in calcium intake (Bryden et al., 2021).

During heat stress, reduced calcium intake and poor absorption result in lower plasma calcium levels, leading to less calcium being available for eggshell formation in laying hens. This results in lower egg output, smaller eggs, or thin-shelled eggs, and poor skeletal development, causing economic losses to producers (Allahverdi et al., 2013; Ventura and Matias da Silva, 2019). As poultry's DM intake is already low due to heat stress, adding large amounts of calcium supplements may not be viable. However, a larger particle-size calcium source including limestone or oyster shells is retained in the gizzard for a longer period and is released slowly into the duodenum for eventual absorption into circulation (Mir et al., 2018).

Electrolytes and vitamins

The main causes of poor performance in heatstressed chickens have been identified by the alteration of the acid-base balance and lowered feed intake (Sugiharto et al., 2017). The minerals potassium (K), sodium (Na), and chlorine (Cl) are essential for maintaining the acidbase balance of bodily fluids (Popoola et al., 2019). As a result, adding minerals such as ammonium chloride (NH₄Cl), sodium bicarbonate (NaHCO₃), sodium chloride (NaCl), potassium chloride (KCl), and potassium sulfate (K₂SO₄) to the diet or drinking water of heat-stressed chickens will assist to mitigate the negative effects of heat stress (Diarra and Tabuaciri, 2014; Pawar et al., 2016).

At high ambient temperatures water intake increases while feed intake decreases. Chickens drink four times more at 38°C compared to 21°C (Orakpoghenor et al., 2020), indicating that water must be available all the time during this period. Increased water intake, which improves heat dissipation and cools down the body provides relief from the detrimental effects of heat exhaustion by supplementing the drinking water with Na+, K+, and Cl⁻ salts (Gamba et al., 2015). Bryden et al. (2021) found that heat-stressed laying hens treated with 0.5% hydrochloric acid in drinking water had significant gains in egg production and egg quality. Gamba et al. (2015) observed that excreta and litter moisture rise due to increased water intake caused by elevated Na⁺ and K⁺ levels.

In another study, Cherian (2015) found that supplementing drinking water with vitamins A, D, E, and B complex increased broiler performance and immune function. Additionally, supplementation of vitamin C (ascorbic acid) has been found to improve performance through improved feed consumption and nutritional intake in heat-stressed birds. Furthermore, Asensio et al. (2020) observed that supplementing broilers with ascorbic acid enhanced the weight and protein content of the carcass while lowering carcass fat content. Daghir (2009) recommended 1 g of vitamin C per liter of drinking water and 20 mg per liter of water for broilers and laying hens, respectively. A study by Wang et al. (2011) in laying hens found that vitamin C does not affect egg weight or egg production. However, Skřivan et al. (2013) reported that 50 and 100 mg/kg vitamin C supplementation significantly increased fertility and hen-day egg production of broiler breeders.

Since poultry cannot synthesize vitamin E, they must be supplemented (Attia et al., 2016). The hormone levels of catecholamine and corticosterone rise in response to stress, particularly heat stress, and lipid peroxidation in cell membranes begins (Abd El-Hack et al., 2018). Vitamin E has also been proven to safeguard macrophages, lymphocytes, and plasma cells from oxidative stress while also enhancing their viability, propagation, and functionality (Shakeri et al., 2020). Therefore, supplementing with vitamin E in the diet during times of stress improves the immunological response of poultry. According to new research, adding vitamin E at a dosage of 250 mg/kg to broiler chickens is a viable protective approach for reducing the severity of heat stress and it may result in optimal performance and enhanced meat quality (Saeed et al., 2019). For layers, however, the dosage is 125-250 mg/kg has been found to result in an improved immunological response, egg production, and feed utilization (Shakeri et al., 2020). Heat stress raises the levels of malondialdehyde in the blood and liver, whereas vitamin E inhibits the formation of malondialdehyde in the liver by preventing lipid peroxidation and cell damage (McDowell, 2012), resulting in improved chicken performance.

Supply of cool water

Water consumption and balance are linked to evaporative heat dissipation and calories dissipated every breath (Chikumba and Chimonyo, 2013; Abdel-Moneim et al., 2021). Reduced water temperature encourages water consumption, which increases evaporative cooling and heat dissipation for each breath (McCreery, 2015). Furthermore, a 20% water consumption increase can result in a 30% increase in heat loss in each breath, with a corresponding performance improvement (Abdel-Moneim et al., 2021).

Water temperature, height, and the shape of drinkers affect poultry performance during heat stress (Orakpoghenor et al., 2020). Water consumption is high in nipple drinkers that are slightly above chick eye than in lower nipple drinkers as chickens find it difficult to bend down and drink from lower nipples (Quilumba et al., 2015; Ranjan et al., 2019). Daghir (2009) recommended the use of wider and deeper drinkers during heat stress as they will permit immersion of not only the beak but the whole face and help dissipate more heat. Cool water at 10-12°C is helpful to poultry, therefore, there is a need to protect water tanks and pipes from the direct sun because birds will not drink warm water (Park et al., 2015). Poultry should always have access to cool, clean water that is below 25°C and has ice in it so that their body temperatures can remain steady during times of heat stress (Park et al., 2015).

Use of phytochemicals in mitigating heat stress

To reduce heat stress in poultry, various phytochemical supplements have been added to the diet.

Resveratrol

Natural bioactive polyphenols called resveratrol are mostly found in peanuts, grapes, turmeric, and berries (Saeed et al., 2017). Resveratrol supplementation (400 mg/kg of feed) has been found in previous studies to boost the antioxidant capacity in broilers under heat stress (Hu et al., 2019). In yellow-feather broilers under heat stress, resveratrol supplementation at 300 or 500 mg/kg of feed daily growth, decreased rectal increased average decreased temperature, and the levels of adrenocorticotropin hormone, malondialdehyde (MDA), corticosterone, and cholesterol (He et al., 2019). Resveratrol supplementation of 200 mg/kg of feed increased egg production in laying hens, whereas resveratrol supplementation of 400 mg/kg of feed decreased total blood cholesterol and triglycerides, decreased egg cholesterol content, increased antioxidant activity, and increased egg sensory scores (Zhang et al., 2017).

Lycopene

The carotenoid lycopene, which is mostly present in tomatoes and tomato-based products, is known to increase the synthesis of antioxidant enzymes by activating the DNA's antioxidant response element (Wasti et al., 2020). Heat-stressed broilers' total feed intake, weight gain, and FCR were all improved when lycopene (200 or 400 mg/kg of feed) was added (Sahin et al., 2016). Lycopene has been reported to increase the levels of antioxidant enzymes, such as superoxide dismutase (SOD) and glutathione peroxidase (GSH-Px) in broilers (Arain et al., 2018). Lycopene administration increased vitamin levels, improved oxidative stability, and the yolk color of eggs in laying hens (Sahin et al., 2016; Arain et al., 2018).

Epigallocatechin gallate

Green tea extract contains the polyphenol epigallocatechin gallate (EGCG), which has strong antiinflammatory and antioxidant effects (Hu et al., 2019). When Luo et al. (2018) fed heat-stressed broiler birds at three EGCG dosages (0, 300, and 600 mg/kg), they observed a linear increase in feed intake, body weight, and levels of blood total protein, glucose, and alkaline phosphatase activity. In a related study, Xue et al. (2017) found that feeding EGCG improved body weight and antioxidant enzyme levels (catalase, GSH-Px, and SOD) in heat-stressed broiler chicks' liver and serum.

Curcumin

The main polyphenols extracted from turmeric are called curcumin, which has anti-inflammatory and antioxidant properties (Attia et al., 2017; Wasti et al., 2020). Even though curcumin is easily absorbed by animals, more recent studies have concentrated on its potential application as a compound to reduce heat stress in chickens (Wasti et al., 2020). It was reported that adding curcumin to feed at a rate of 100 mg/kg significantly increased broiler body weight during heat stress (Zhang et al., 2017). Furthermore, the inclusion of 150 mg/kg of curcumin in the diet of laying hens enhanced egg quality, laying efficiency, antioxidant enzyme activity, and immunological response to heat stress (Liu et al., 2020).

Mitigation of heat stress by use of probiotics and betaine

Betaine is widely distributed in plants, animals, microbes, and its rich food sources include fish, spinach, and wheat bran (Saeed et al., 2017). Betaine plays an essential role in sustaining the basic functions of poultry, including osmoregulation, fat distribution, methionine sparing, immunity, and the bird's ability to withstand heat stress (Attia et al., 2016; Saeed et al., 2019). The performance of chickens kept under heat stress can be greatly improved by including betaine in their diets (Hao et al., 2017; Saeed et al., 2017). In addition, betaine also functions as a methyl donor, which enables feed cost reductions by substituting methionine and choline supplements (Gholami et al., 2015). Betaine supports a variety of intestinal bacteria in their defense against osmotic changes, improving microbial fermentation activity (Abd El-Ghany and Babazadeh, 2022).

The term "probiotics" refers to feed additives that contain live beneficial microorganisms such as Bifidobacterium, Streptococci, and Lactobacillus, yeast cultures with Saccharomyces and candida strains, and fungi (*Aspergillus awamori*, *A. niger*, and *A. oryza*), which may improve poultry performance, intestinal microbiota, and immune system (Abd El-Hack, et al., 2018; El-Moneim et al., 2020). Probiotics have received a lot of attention lately for reducing the oxidative damage brought about by heat stress in chickens (Ahmad et al., 2022). It has been shown that the addition of probiotics to the diet of broilers increased their growth performance, FCR, and immunological response (Wang et al., 2018).

A symbiotic relationship occurs when prebiotics and probiotics are combined to have a positive effect on poultry raised in hot environments (Lara and Rostagno, 2013). It has been suggested that incorporating synbiotics in the diet may benefit chickens kept in areas that experience high levels of heat stress by minimizing the negative effects of heat and possibly improving their welfare and performance (Mohammed et al., 2018). Probiotic supplements have been shown to have favorable benefits on the health and productivity of chickens in tropical climates (Ahossi et al., 2016; Deraz, 2018). It was reported that the performance, intestinal morphology, and immunological response of heatstressed chickens were all improved by consuming mannanoligosaccharides, prebiotics, and a probiotic combination (Jahromi et al., 2015).

CONCLUSION

Heat stress has a negative impact on the health and productivity of poultry and is a significant challenge in poultry production in the tropics. Heat stress results from a combination of many factors including high ambient temperature, radiant heat, humidity, and airspeed. Due to heat stress many behavioral, neuroendocrinal, and physiological changes occur. Gene screening for higher growth rates to meet the ever-increasing food requirement has made poultry susceptible to heat stress. In birds raised for egg and meat production, an increase in the ambient temperature induces decreases in body weight gain, feed intake, eggshell weight, higher FCR, and increases in body temperature. These negative effects can be addressed by strategic managerial enhancements. Several approaches are used worldwide to combat the severe impacts of heat stress, including the selection of rearing systems with better ventilation, suitable housing conditions, and recommended correct stocking densities, all of which are crucial for enhancing performance at high temperatures.

Given that there is no single strategy for heat stress, a variety of strategies will help to reduce it. Further research on new innovative strategies which include utilizing heat tolerance genes and selecting genotypes with higher heat tolerance using genetic markers should be carried out.

DECLARATIONS

Acknowledgments

The researchers would like to thank the reviewers for their constructive criticisms that helped to improve the quality of this manuscript.

Funding

None.

Authors' contributions

Shame Bhawa and John Cassius Moreki conceptualized this study. Shame Bhawa surveyed the literature, and drafted and revised the manuscript while John Moreki edited and suggested changes to the manuscript. James Butti Machete also surveyed and played a part in drafting the manuscript. All authors checked and approved the final version of the manuscript for publication in this journal.

Competing interests

The authors declare no existence of competing for interests.

Ethical considerations

The authors have examined ethical issues, such as plagiarism, permissions to publish, misconduct, and duplicate publishing.

REFERENCES

- Abbas G, Arshad M, Tanveer A, Jabbar M, Mahmood A, Khan M, Konca Y, Sultan Z, Qureshi R, Iqbal A et al. (2021). Combating heat stress in laying hens a review. Pakistan Journal of Science, 73(4): 633-655. Available at: <u>http://pjosr.com/index.php/pjs/article/view/365/211</u>
- Abd El-Ghany WA and Babazadeh D (2022). Betaine: A potential nutritional metabolite in the poultry industry. Animals, 12(19): 2624. DOI: <u>https://www.doi.org/10.3390/ani12192624</u>
- Abd El-Hack A, Mohamed E, Alagawany M, and Noreldin AE (2018). Managerial and nutritional trends to mitigate heat stress risks in poultry farms. In: A. Negm and M. Abu-hashim (Editros). Sustainability of agricultural environment in Egypt: Part II. The handbook of environmental chemistry. Springer., Cham. 77: 325-338. DOI: <u>https://www.doi.org/10.1007/698_2018_290</u>
- Abdel-Moneim AE, Shehata AM, Khidr RE, Paswan VK, Ibrahim NS, El-Ghoul AA, Aldhumri SA, Gabr SA, Mesalam NM, Elbaz AM et al. (2021). Nutritional manipulation to combat heat stress in poultry A comprehensive review. Journal of Thermal Biology, 98: 102915. DOI: https://www.doi.org/10.1016/j.jtherbio.2021.102915
- Adu-Asiamah P, Zhang Y, Amoah K, Leng QY, Zheng JH, Yang H, Zhang WL, and Zhang L (2021). Evaluation of physiological and

molecular responses to acute heat stress in two chicken breeds. Animal, 15(2): 100106. DOI: https://www.doi.org/10.1016/j.animal.2020.100106

- Afsal A, Sejian V, Bagath M, Devaraj C, and Bhatta R (2018). Heat stress and livestock adaptation: Neuro-endocrine regulation. International Journal of Veterinary and Animal Medicine, 1(2): 1-7. DOI: <u>https://www.doi.org/10.31021/ijvam.20181108</u>
- Afsharmanesh M, Lofti M, and Mehdipour Z (2016). Effects of wet feeding and early feed restriction on blood parameters and growth performance of broiler chickens. Animal Nutrition, 2(3): 168-172. DOI: <u>https://www.doi.org/10.1016/j.aninu.2016.04.002</u>
- Ahmad R, Yu YH, Hsiao FSH, Su CH, Liu HC, Tobin I, Zhang G, and Cheng YH (2022). Influence of heat stress on poultry growth performance, intestinal inflammation, and immune function and potential mitigation by probiotics. Animals, 12(7): 2297. DOI: https://www.doi.org/10.3390/ani12172297
- Ahmed ME and Abbas TE (2013). The effect of feeding pellets versus mash on performance and carcass characteristics of broiler chicks. Bulletin of Environment, Pharmacology and Life Sciences, 2(2): 31-34. Available at: <u>https://bepls.com/jan_2013/8.pdf</u>
- Ahossi PK, Dougnon JT, Kiki PS, and Houessionon JM (2016). Effects of Tridax procumbens powder on zootechnical, biochemical parameters and carcass characteristics of Hubbard broiler chicken. Journal of Animal Health and Production, 4(1): 15-21. Available at: http://www.nexusacademicpublishers.com/uploads/files/JAHP_MH2 0151110121122_%20Ahossi%20et%20al.pdf
- Akbarian A, Michiels J, Degroote J, Majdeddin M, Golian A, and De Smet S (2016). Association between heat stress and oxidative stress in poultry; mitochondrial dysfunctionand dietary interventions with phytochemicals. Journal of Animal Science and Biotechnology, 7: 37. Available at: <u>https://jasbsci.biomedcentral.com/articles/10.1186/s40104-016-0097-5?report=reader</u>
- Alchalabi AD (2013). Poultry housing design. Chapter 3. Pp. 1-8. DOI: https://www.doi.org/10.13140/2.1.2729.7280
- Allahverdi A, Feizi A, Takhtfooladi HA, and Nikpiran H (2013). Effects of heat stress on acid-base imbalance, plasma calcium concentration, egg production and egg quality in commercial layers. Global Veterinaria, 10(2): 203-207.
- Altan O, Pabuccuoglu A, Altan A, Konyalioglu S, and Bayraktar H (2003). Effect of heat stress on oxidative stress, lipid peroxidation and some stress parameters in broilers. British Poultry Science, 44: 545-550. DOI: doi:<u>https://www.doi.org/10.1080/00071660310001618334</u>

Alverdy JC and Luo JN (2017). The influence of host stress on the mechanism of infection: Lost microbiomes, emergent pathobiomes, and the role of interkingdom signalling. Frontiers in Microbiology, 8: 322. DOI: <u>https://www.doi.org/10.3389/fmicb.2017.00322</u>

- Arain MA, Mei Z, Hassan FU, Saeed M, Alagawany M, Shar AH, and Rajput IR (2018). Lycopene: A natural antioxidant for prevention of heat-induced oxidative stress in poultry. World's Poultry Science Journal, 74(1): 89-100. DOI: https://www.doi.org/10.1017/S0043933917001040
- Archana PR, Aleena J, Pragna P, Vidya MK, Abdul Niyas PA, Bagath M, Krishnan G, Manimaran A, Beena V, Kurien EK et al. (2017). Role of heat shock proteins in livestock adaptation to heat stress. Journal of Dairy Veterinary and Animal Research, 5(1): 13-19. DOI: https://www.doi.org/10.15406/jdvar.2017.05.00127
- Arnal M E and Lallès JP (2016). Gut epithelial inducible heat-shock proteins and their modulation by diet and the microbiota. Nutrition Reviews, 74(3): 181-197. DOI: https://www.doi.org/10.1093/nutrit/nuv104
- Asensio X, Abdelli N, Piedrafita J, Soler MD, and Barroeta AC (2020). Effect of fibrous diet and vitamin C inclusion on uniformity, carcass traits, skeletal strength, and behaviour of broiler breeder pullets. Poultry Science, 99(5): 2633-2644. DOI: https://www.doi.org/10.1016/j.psj.2020.01.015
- Aswathi PB, Bhanja SK, Kumar P, Shyamkumar TS, Mehra M, Bhaisare DB, and Rath PK (2019). Effect of acute heat stress on the physiological and reproductive parameters of broiler breeder hens–A study under controlled thermal stress. Indian Journal of Animal Research, 53(9): 1150-1155. Available at:

https://arccjournals.com/journal/indian-journal-of-animal-research/B-3641

- Attia YA, Abd El-Hamid AEHE, Abedalla AA, Berika MA, Al-Harthi MA, Kucuk O, Sahin K, and Abou-Shehema BM (2016). Laying performance, digestibility and plasma hormones in laying hens exposed to chronic heat stress as an ected by betaine, vitamin C, and/or vitamin E supplementation. SpringerPlus, 5: 1619. DOI: https://www.doi.org/10.1186%2Fs40064-016-3304-0
- Attia YA, Al-Harthi MA, and Hassan SS (2017). Turmeric (Curcuma longa Linn.) as a phytogenic growth promoter alternative for antibiotic and comparable to mannan oligosaccharides for broiler chicks. Revista Mexicana de Ciencias Pecuarias, 8: 11-21. DOI: <u>https://www.doi.org/10.22319/rmcp.v8i1.4309</u>
- Attia YA, Bovera F, Wang J, Al-Harthi MA, and Kim WK (2020). Multiple amino acid supplementations to low- protein diets: Effect on performance, carcass yield, meat quality and nitrogen excretion of g finishing broilers under hot climate conditions. Animals, 10(6): 973. DOI: <u>https://www.doi.org/10.3390/ani10060973</u>
- Awad E, Zulkifli I, Soleimani A, Law F, Ramiah S, Yousif MIM, Hussein EA, and Khalil ES (2019). Response of broilers to reduced protein diets under heat stress conditions. World's Poultry Science Journal, 75(4): 583-598. DOI: https://www.doi.org/10.1017/S0043933919000576
- Azhar M, Mahmud A, Usman M, Javed K, Ishaq H, Mehmood S, Ahmad S, Hussain J, Ghayas A, and Abbas M (2019). Effect of breeder age on the progeny performance of three naked-neck chicken phenotypes. Brazilian Journal of Poultry Science, 21(3): 1-6. DOI: https://www.doi.org/10.1590/1806-9061-2018-0729
- Babazadeh D and Ahmadi Simab P (2022). Methionine in poultry nutrition: A review. Journal of World's Poultry Science, 1(1): 1-11. Available at: https://jwps.rovedar.com/article_138282.html
- Badruzzaman ATM, Noor M, AL Mamun M, Husna A, Islam KM, and Rahman MM (2015). Prevalence of diseases in commercial chickens at Sylhet division of Bangladesh. International Clinical Pathology Journal, 1(5): 104-108. DOI: https://www.doi.org/10.15406/icpj1.2015.01.00023
- Barrett NW, Rowland K, Schmidt CJ, Lamont SJ, Rothschild MF, Ashwell CM, and Persia ME (2019). Effects of acute and chronic heat stress on the performance, egg quality, body temperature, and blood gas parameters of laying hens. Poultry Science, 98(12): 6684-6692. DOI: <u>https://www.doi.org/10.3382/ps/pez541</u>
- Beckford RC, Ellestad LE, Weglarz MP, Farley L, Brady K, Angel R, Liu HC, and Porter TE (2020). Effects of heat stress on performance, blood chemistry, and hypothalamic and pituitary mRNA expression in broiler chickens. Poultry Science, 99(12): 6317-6325. DOI: https://www.doi.org/10.1016/j.psj.2020.09.052
- Bernabucci U (2019). Climate change: Impact on livestock and how can we adapt. Animal Frontiers, 9(1): 3-5. DOI: https://www.doi.org/10.1093/af/vfz022
- Bhadauria P (2017). Different types of poultry housing system for tropical climate. Indian Council of Agricultural Research, pp. 1-68.
- Bhadauria P, Keshava MP, Murai A, and Jadoun YS (2016). Management of heat stress in poultry production system. ICAR Agricultural Technology Application Research Institute, Zone-1, Ludhiana 141: 4.
- Bilal RM, Hassan F, Farag RM, Nasir TA, Ragni M, Mahgoub HAM, and Alagawany M (2021). Thermal stress and high stocking densities in poultry farms: Potential effects and mitigation strategies. Journal of Thermal Biology, 99: 102944. DOI: https://www.doi.org/10.1016/j.jtherbio.2021.102944
- Bohler MW, Chowdhury VS, Cline MA, and Gilbert ERR (2021). Heat stress responses in birds: A review of the neural components. Biology, 10(11): 1095. DOI: https://www.doi.org/10.3390/biology10111095
- Bryden WL, Li X, Ruhnke I, Zhang D, and Shini S (2021). Nutrition, feeding and laying hen welfare. Animal Production Science, 61(10): 893-914. DOI: <u>https://www.doi.org/10.1071/AN20396</u>
- Cadirci S and Koncagul S (2014). Possible effects of delivering methionine to broilers in drinking water at constant low and high environmental temperatures. Italian Journal of Animal Science, 13(1): 3013. DOI: https://www.doi.org/10.4081/ijas.2014.3013

https://www.doi.org/10.1016/j.vetimm.2016.02.004

- Calefi AS, Honda BT, Costola-de-Souza C, de Siqueira A, Namazu LB, Quinteiro-Filho WM, Fonseca JG, Aloia TP, Ferreira AJP, and Palermo-Neto J (2014). Effects of long term heat stress in an experimental model of avian necrotic enteritis. Poultry Science, 93(6): 1344-1353. DOI: <u>https://www.doi.org/10.3382/ps.2013-03829</u>
- Campbell ZA, Otieno L, Shirima GM, Marsh TL, and Palmer GH (2019). Drivers of vaccination preferences to protect a low-value livestock resource: Willingness to pay for Newcastle disease vaccines by smallholder households. Vaccine, 37(1): 11-8. DOI: https://www.doi.org/10.1016/j.vaccine.2018.11.058
- Cantet JM, Yu Z, and Ríus AG (2021). Heat stress- mediated activation of immune–inflammatory pathways. Antibiotic, 10(11): 1285. DOI: <u>https://www.doi.org/10.3390/antibiotics10111285</u>
- Cao C, Chowdhury VS, Cline MA, and Gilbert ER (2021). The microbiota-gut-brain axis during heat stress in chickens: A review. Frontiers in Physiology, 12: 752265. DOI: https://www.doi.ord/10.3389/fphys.2021.752265
- Cassuce D, Tinôco I, Baêta F, Zolnier S, Cecon P, and Vieira MF (2013). Thermal comfort temperature update for broiler chickens up to 21 days of age. Engenharia Agrícola, 33(1): 28-36. DOI: https://www.doi.org/10.1590/S0100-69162013000100004
- Cedraz H, Gromboni JGG, Garcia AAP Junior, Farias Filho RV, Souza TM, de Oliveira ER, de Oliverira EB, do uscimento ES, Meneghetti C, and Wanceslau AA (2017). Heat stress induces expression of HSP genes in genetically divergent chickens. PloS ONE, 12(10): e0186083. DOI: https://www.doi.org/10.1371/journal.pone.0186083
- Cherian G (2015). Nutrition and metabolism in poultry: Role of lipids in the early diet. Journal of Animal Science and Biotechnology, 6: 28. DOI: <u>https://www.doi.org/10.1186/s40104-015-0029-9</u>
- Chikumba N and Chimonyo M (2013). Effects of water restriction on the growth performance, carcass characteristics and organ weights of Naked Neck and Ovambo chickens of Southern Africa. Asian-Australasian Journal of Animal Sciences, 27(7): 974-980. DOI: https://www.doi.org/10.5713%2Fajas.2013.13383
- Cioffi F, Senese R, Lanni A, and Goglia F (2013). Thyroid hormones and mitochondria: with a brief look at derivatives and analogues. Molecullar and Cellular Endocrinology, 379(1-2): 51-61. DOI: https://www.doi.org/10.1016/j.mce.2013
- Daghir NJ (2009). Nutritional strategies to reduce heat stress in broilers and broiler breeders. Lohmann Information, 44(1): 6-15. Available at: http://www.lohmann-information.com/co Accessed 19/10/2020
- Dao HT, Sharma NK, Bradbury EJ, and Swick RA (2021). Effects of Larginine and L-citrulline supplementation in reduced protein diets for broilers under normal and cyclic warm temperature. Animal Nutrition, 7(4): 927-938. DOI: https://www.doi.org/10.1016/j.aninu.2020.12.010
- Datta K, Rahalkar K, and Dinesh DK (2017). Heat shock proteins (HSP): Classifications and its involvement in health and disease. Journal of Pharmaceutical Care & Health Systems, 4(2): 1000175. DOI: https://www.doi.org/10.4172/2376-0419.1000175
- Dayyani N and Bakhtiyari H (2013). Heat stress in poultry: Background and affective factors. International Journal of Advanced Biological and Biomedical Research, 1(11): 1409-1413. Available at: http://www.ijabbr.com/article_7968_f56af19891048da54f54040274e 4cd73.pdf
- De Almeida A, Dias A, Bueno CFD, Couto FAP, Rodrigues PA, Nogueira WCL, and Emygdio de Faria Filho D (2012). Crude protein and metabolizable energy levels for layers reared in hot climates. Revista Brasileira de Ciência Avícola, 14: 203-208. DOI: https://www.doi.org/10.1590/S1516-635X2012000300007
- de Freitas Dionizio A, de Souza Khatlab A, Alcalde CR, Gasparino E, and Feihrmann AC (2021). Supplementation with free methionine or methionine dipeptide improves meat quality in broilers exposed to

heat stress. Journal of Food Science and Technology, 58(1): 205-215. DOI: https://www.doi.org/10.1007/s13197-020-04530-2

- De Oliveira RG and Lara LJC (2016). Lighting programmes and its implications for broiler chickens. World's Poultry Science Journal, 72(4): 735-742. DOI: https://www.doi.org/10.1017/S0043933916000702
- Del Vesco A, Gasparino PE, Neto ARO, Rossi RM, Soares MAM, and da Silva SCC (2013). Effect of Methionine supplementation on mitochondrial genes expression in the breast muscle and liver of broilers. Livestock Science, 151(2-3): 284-291. Available at: https://www.infona.pl/resource/bwmeta1.element.elsevier-9c387820-6b46-316a-a936-58722a56f0cb
- Deraz SF (2018). Synergetic effects of multispecies probiotic supplementation on certain blood parameters and serum biochemical profile of broiler chickens. Journal of Animal Health and Production, 6(1): 27-34. DOI: https://www.doi.org/10.17582/journal.jahp/2018/6.1.27.34
- Desta TT (2021). The genetic basis and robustness of naked neck mutation in chicken. Tropical Animal Health and Production, 53: 95.
- DOI: <u>https://www.doi.org/10.1007/s11250-020-02505-1</u> Diarra SS and Tabuaciri P (2014). Feeding management of poultry in high environmental temperatures. International Journal of Poultry
- Science, 13(11): 657-661. DOI: https://www.doi.org/10.3923/ijps.2014.657.661 Diarra SS, Sandakabatu D, Perera D, Tabuaciri P, and Mohammed U
- Diarra SS, Sandakabatu D, Perera D, Tabuaciri P, and Mohammed U (2014). Growth performance and carcass yield of broiler chickens fed commercial finisher and cassava copra meal-based diets. Journal of Applied Animal Research, 43(3): 352-356. DOI: <u>https://www.doi.org/10.1080/09712119.2014.978774</u>
- Donald J (2018). How poultry housing can optimise performance. Biomin, Auburn University, in Alabama, USA. https://www2.biomin.net/hr/articles/how-poultry-housing-canreduce-stress-and-optimize-performance/
- Dong J, Chuan H, Zhibing W, Yanqing L, Shanshan L, Lin T, Jiebo C, Donghua, L, Fenxia Y, Naibin L et al. (2018). A novel deletion in KRT75L4 mediates the frizzle trait in a Chinese indigenous chicken. Genetics Selection Evolution, 50: 68. DOI: https://www.doi.org/10.1186/s12711-018-0441-7
- Duangjinda MI, Tunim S, Duangdaen CI, and Boonkum W (2017). HSP70 genotypes and heat tolerance of commercial and native chickens reared in hot and humid conditions. Brazilian Journal of Poultry Science, 19(1): 7-18. DOI: http://www.doi.org/10.1590/1806-9061-2016-024
- Ebeid TA, Suzuki T, and Sugiyama T (2012). High ambient temperature influences eggshell quality and calbindin D28k localization of eggshell gland and all intestinal segments of laying hens. Poultry Science, 91(9): 2282-2287. DOI: https://www.doi.org/10.3382/ps.2011-01898
- Efeoğlu B (2009). Heat shock proteins and heat shock response in plants. Ghazi University Journal of Science, 22(2): 67-75. Available at: https://www.acarindex.com/pdfler/acarindex-43ec2602-cd1b.pdf
- Elbaz AM, Ibrahim NS, Shehata AM, Mohamed NG, and Abdel-Moneim A-ME (2021). Impact of multi-strain probiotic, citric acid, garlic powder or their combinations on performance, ileal histomorphometry, microbial enumeration and humoral immunity of broiler chickens. Tropical Animal Health and Production, 53: 115. DOI: <u>https://www.doi.org/10.1007/s11250-021-02554-0</u>
- Estévez M (2015). Oxidative damage to poultry: From farm to fork. Poultry Science, 94(6): 1368-1378. DOI: https://www.doi.org/10.3382/ps/pev094
- Farag MR and Alagawany M (2018). Physiological alterations of poultry to the high environmental temperature. Journal of Thermal Biology, 76(1): 101-106. DOI: https://www.doi.org/10.1016/j.jtherbio.2018.07.012
- Farghly MFA, Abd El-Hack ME, Alagawany M, Saadeldin IM, and Swelum AA (2018). Wet feed and cold water as heat stress modulators in growing Muscovy ducklings. Poultry Science, 97(5): 1588-1594. DOI: <u>https://www.doi.org/10.3382/ps/pey006</u>
- Fathi M, Al-Homidan I, Rayan G, El-Safty S, Ebeid T, and Abou-Emera O (2019). Laying performance, immune response and antioxidant properties of hens segregating for naked neck and frizzle genes under

low ambient temperature. Czech Journal of Animal Science, 64(5): 216-225. DOI: http://www.doi.org/10.17221/221/2018-CJAS

- Fathi M, Galal AA, El-Saft SAE, and Mahrous M (2013). Naked neck and frizzle genes for improving chickens raised under high ambient temperature: I. Growth performance and egg production. World's Poultry Science Journal, 69(4): 813-832. DOI: http://www.doi.org/10.1017/S0043933913000834
- Fathi MM, Galal A, Radwan LM, Abou-Emera OK, and Al-Homidan IH (2022). Using major genes to mitigate the deleterious effects of heat stress in poultry: An updated review. Poultry Science, 101(11): 102157. DOI: <u>https://www.doi.org/10.1016/j.psj.2022.102157</u>
- Faud AM, Chen W, Ruan D, Wang S, Xia WG, and Zheng CT (2016). Impact of heat stress on meat, egg quality, immunity and fertility in poultry and nutritional factors that overcome these effects: A review. International Journal of Poultry Science, 15(3): 81-95. DOI: http://www.doi.org/10.3923/ijps.2016.81.95
- Gamba JP, Rodrigues MM, Garcia Neto M, Perri SH, Faria de Júnior MJA, and Pinto MF (2015). The strategic application of electrolyte balance to minimize heat stress in broilers. Brazilian Journal of Poultry Science, 17(2): 237-246. DOI: http://www.doi.org/10.1590/1516-635x1702237-246
- Gholami J, Qotbi AAA, Seidavi A, Meluzzi A, Tavaniello S, and Maiorano G (2015). Effects of *in ovo* administration of betaine and choline on hatchability results, growth and carcass characteristics and immune response of broiler chickens. Italian Journal of Animal Science, 14(2): 3694. DOI: https://www.doi.org/10.4081/ijas.2015.3694
- Gholami M, Chamani M, Seidavi A, Asgha S, and Aminafshar M (2020). Effects of stocking density and climate region on performance, immunity, carcass characteristics, blood constituency, and economical parameters of broilers. Revista Brasileira de Zootecnia, 49: e20190049. DOI: https://www.doi.org/10.37496/rbz4920190049
- Gicheha MG (2021). The effects of heat stress on production, reproduction, health in chicken and its dietary amelioration. In (Ed.), Advances in poultry research, IntechOpen. DOI: https://www.doi.org/10.5772/intechopen.97284
- Glatz P and Pym R (2013). Poultry Development Review, poultry housing and management in developing countries. FAO., Rome, Italy. pp. 24-28. Available at: https://www.fao.org/3/i3531e/i3531e.pdf#page=30
- Goel A (2021). Heat stress management in poultry. Journal of Animal Physiology and Animal Nutrition, 105(6): 1136-1145. DOI: https://www.doi.org/10.1111/jpn.13496
- Gonzalez-Rivas PA, Chauhan SS, Ha M, Fegan N, Dunshea FR, and Warner RD (2020). Effects of heat stress on animal physiology, metabolism, and meat quality: A review. Meat Science, 162: 108025. DOI: <u>https://www.doi.org/10.1016/j.meatsci.2019.108025</u>
- Habashy WS, Milfort MC, Fuller AL, Attia YA, Rekaya R, and Aggrey SE (2017). Effect of heat stress on protein utilization and nutrient transporters in meat-type chickens. International Journal of Biometeorology, 61: 2111-2118. DOI: https://www.doi.org/10.1007/s00484-017-1414-1
- Hamm LL, Nakhoul N, and Hering-Smith KS (2015). Acid-Base base homeostasis. Clinical journal of the American Society of Nephrology, 10(12): 2232-2242. DOI: https://www.doi.org/10.2215/CJN.07400715
- Hao S, Liu L, Wang G, Xian GU, and Pan F (2017). Effects of dietary betaine on performance, egg quality and serum biochemical parameters of laying hens under heat stress condition. Chinese Journal of Animal Nutrition, 29(1): 184-192. Available at: http://www.chinajan.com/CN/abstract/html/20170121.htm
- He S, Li S, Arowolo MA, Yu Q, Chen F, Hu R, and He J (2019). Effect of resveratrol on growth performance, rectal temperature, and serum parameters of yellow-feather broilers under heat stress. Animal Science Journal, 90(3): 401-411. DOI: https://www.doi.org/10.1111/asj.13161
- Herman JP, McKlveen JM, Ghosal S, Kopp B, Wulsin A, Makinson R, Scheimann J, and Myers B (2016). Regulation of the hypothalamicpituitary- adrenocortical stress response. Comprehensive Physiology, 6(2): 603-621. DOI: <u>https://www.doi.org/10.1002/cphy.c150015</u>

- Hirakawa R, Nurjanah S, Furukawa K, Murai A, Kikusato M, Nochi T, and Toyomizu M (2020). Heat stress causes limmune abnormalities via massive damage to effect proliferation and differentiation of lymphocytes in broiler chickens. Frontiers in Veterinary Science, 7: 46. DOI: <u>https://www.doi.org/10.3389/fvets.2020.00046</u>
- Holtmann G, Shah A, and Morrison M (2017). Pathophysiology of functional gastrointestinal disorders: A holistic overview. Digestive Diseases, 35(Suppl 1): 5-13. DOI: https://www.doi.org/10.1159/000485409
- Hu JY, Hester PY, Makagon MM, Vezzoli G, Gates RS, Xiong YJ, and Cheng HW (2016). Cooled perch effects on performance and wellbeing traits in caged white leghorn hens. Poultry Science, 95(12): 2737-2746. DOI: <u>https://www.doi.org/10.3382/ps/pew248</u>
- Hu R, He Y, Arowolo MA, Wu S, and He J (2019). Polyphenols as potential attenuators of heat stress in poultry production. Antioxidants, 8(3): 67. DOI: <u>https://www.doi.org/10.3390/antiox8030067</u>
- Jahanian R and Rasouli E (2015). Dietary chromium methionine supplementation could alleviate immunosuppressive effects of heat stress in broiler chicks. Journal of Animal Science, 93(7): 3355-3363. DOI: <u>https://www.doi.org/10.2527/jas.2014-8807</u>
- Jahromi MF, Altaher YW, Shokryazdan P, Ebrahimi R, Ebrahimi M, Idrus Z, Tufarlli V, and Lian JB (2015). Dietary supplementation of a mixture of Lactobacillus strains enhances performance of broiler chickens raised under heat stress conditions. International Journal of Biometeorology, 60(7): 1099-1110. DOI: https://www.doi.org/10.1007/s00484-015-1103-x
- Jessop TS, Meagan LL, Teasdale L, Stuart-Fox D, Willson RS Careau V, and Moore IT (2016). Multiscale evaluation of thermal dependence in the glucocorticoid response of vertebrates. The American Naturalist, 188(3): 342-356. DOI: https://www.doi.org/10.1086/687588
- Jha R and Mishra P (2021). Dietary fibre in poultry nutrition and their effects on nutrient utilisation, performance, gut health, and on the environment: A review. Journal of Animal Science and Biotechnology, 12: 51. DOI: <u>https://www.doi.org/10.1186/s40104-021-00576-0</u>
- Kadykalo S, Roberts T, Thompson M, Wilson J, Lang M, and Espeisse O (2018). The value of anticoccidials for sustainable global poultry production. International Journal of Antimicrobial Agents, 51(3): 304-310.
 bOI: https://www.doi.org/10.1016/j.ijantimicag.2017.09.004
- Kaldhusdal M, Benestad SL, and Løvland A (2016). Epidemiologic aspects of necrotic enteritis in broiler chickens – disease occurrence and production performance. Avian Pathology, 45(3): 271-274. DOI: https://www.doi.org/10.1080/03079457.2016.1163521
- Kennedy GM, Lichoti KJ, and Ommeh SC (2022). Review article: Heat stress and poultry: Adaptation to climate change, challenges, and opportunities for genetic breeding in Kenya. Journal of Agriculture Science and Technology, 21(1): 49-61. DOI: https://www.doi.org/10.4314/jagst.v21i1.6
- Khalil MM, Abdollahi MR, Zaefarian F, and Ravindran V (2021). Influence of feed form on the apparent metabolisable energy of feed ingredients for broiler chickens. Animal Feed Science and Technology, 271: 114754. DOI: https://www.doi.org/10.1016/j.anifeedsci.2020.114754
- Kidd MT, Maynard CW, and Mullenix GJ (2021). Progress of amino acid nutrition for diet protein reduction in poultry. Journal of Animal Science and Biotechnolology, 12: 45. DOI: https://www.doi.org/10.1186/s40104-021-00568-0
- Kumar BC, Gloridoss RG, Singh KC, Prabhu TM, and Suresh BN (2016). Performance of broiler chickens fed low protein, limiting amino acid supplemented diets formulated either on total or standardized ileal digestible amino acid basis. Asian Australasian Journal of Animal Science, 29(11): 1616-1624. DOI: http://www.doi.org/10.5713/ajas.15.0648
- Kumari NRK and Nath ND (2018). Ameliorative measures to counter heat stress in poultry. World's Poultry Science Journal, 74(1): 117-130. DOI: <u>https://www.doi.org/10.1017/S0043933917001003</u>
- Kumbhar S, Khan AZ, Parveen F, Nizamani ZA, Siyal FA, Mohamed E, Abd El-Hack ME, Gan F, Liu Y, Hamid M et al. (2018). Impacts of

selenium and vitamin E supplementation on mRNA of heat shock proteins, selenoproteins and antioxidants in broilers exposed to high temperature. AMB Express, 8: 112. DOI: https://www.doi.org/10.1186/s13568-018-0641-0

- Lara LJ and Rostagno MH (2013). Impact of heat stress on poultry production. Animals, 3(2): 356-369. DOI: https://www.doi.org/10.3390/ani3020356
- Lemme A, Hiller P, Klahsen M, Taube V, Stegemann J, and Simon I (2019). Reduction of dietary protein in broiler diets not only reduces n-emissions but is also accompanied by several further benefits. Journal of Applied Poultry Research, 28(4): 867-880. DOI: https://www.doi.org/10.3382/japr/pfz045
- Liu F, de Ruyter EM, Athorn RZ, Brewster CJ, Henman DJ, Morrison RS, Smits RJ, Cottrell JJ, and Dunshea FR (2019). Effects of Lcitrulline supplementation on heat stress physiology, lactation performance and subsequent reproductive performance of sows in summer. Journal of Animal Physiology and Animal Nutrition, 103(1): 251-257. DOI: https://www.doi.org/10.1111/jpn.13028
- Liu M, Lu Y, Gao P, Xie X, Li D, Yu D, and Yu M (2020). Effect of curcumin on laying performance, egg quality, endocrine hormones, and immune activity in heat-stressed hens. Poultry Science, 99(4): 2196-2202. DOI: <u>https://www.doi.org/10.1016/j.psj.2019.12.001</u>
- Liverpool-Tasie LSO, Sanou A, and Tambo JA (2019). Climate change adaptation among poultry farmers: Evidence from Nigeria. Climate Change, 157(1): 527-544. DOI: https://www.doi.org/10.1007/s10584-019-02574-8
- Luo J, Song J, Liu L, Xue B, Tian G, and Yang Y (2018). Effect of epigallocatechin gallate on growth performance and serum biochemical metabolites in heat-stressed broilers. Poultry Science, 97(2): 599-606. DOI: <u>https://www.doi.org/10.3382/ps/pex353</u>
- Mack LA, Felver-Gant JN, Dennis RL, and Cheng HW (2013). Genetic variations alter production and behavioural responses following heat stress in 2 strains of laying hens. Poultry Science, 92(2): 285-294. DOI: <u>https://www.doi.org/10.3382/ps.2012-02589</u>
- Massuquetto A, Durau J, Barrilli L, Santos R, Krabbe E, and Maiorka A (2020). Thermal processing of corn and physical form of broiler diets. Poultry Science, 99(6): 3188-3195. DOI: https://www.doi.org/10.1016/j.psj.2020.01.027
- McCreery DH (2015). Water consumption behaviour in broilers. Graduate Theses and Dissertations. Available at: https://scholarworks.uark.edu/etd/1301
- McDowell LR (2012). Vitamins in animal nutrition: Comparative aspects to human nutrition. Elsevier., Amsterdam, The Netherlands.
- McGregor BA, Murphy KM, Albano DL, and Ceballos RM (2016). Stress, cortisol, and B lymphocytes: A novel approach to understanding academic stress and immune function. Stress, 19(2): 185-191. DOI:

https://www.doi.org/10.3109/10253890.2015.1127913

- Mir SH, Pal RP, Mani V, Malik TA, Ojha L, and Yadav S (2018). Role of dietary minerals in heat-stressed poultry: A review. Journal of Entomology and Zoology Studies, 6(5): 820-826.
- Mishra B and Jha R (2019). Oxidative stress in the poultry gut: Potential challenges and interventions. Frontiers in Veterinary Science, 6(1): 60. DOI: <u>http://www.doi.org/10.3389/fvets.2019.00060</u>
- Mohammed AA, Jacobs JA, Murugesan GR, and Cheng HW (2018). Effect of dietary synbiotic supplement on behavioral patterns and growth performance of broiler chickens reared under heat stress. Poultry Science, 97(4): 1101–1108. DOI: https://www.doi.org/10.3382/ps/pex421
- Monson MS, Van Goor AG, Ashwell CM, Persia ME, Rothschild MF, Schmidt CJ, and Lamont SJ (2018). Immunomodulatory effects of heat stress and lipopolysaccharide on the bursal transcriptome in two distinct chicken lines. BMC genomics, 19(1): 643. DOI: https://www.doi.org/10.1186/s12864-018-5033-y
- Moraes PO, Andretta I, Cardinal KM, Ceron M, Vilella L, Borille R, Frazzon AP, Frazzon J, Santin E, and Ribeiro AML (2019). Effect of functional oils on the immune response of broilers challenged with Eimeria spp., Animal, 13(10): 2190-2198. DOI: https://www.doi.org/10.1017/S1751731119000600
- Moreki JC, Magapatona S, and Manyeula F (2020). Effect of stocking density on performance of broiler chickens. International Journal of

Agriculture and Rural Development, 23(2): 5367-5372. Available at: http://researchhub.buan.ac.bw/bitstream/handle/13049/40/IJARD%2 02020.pdf?sequence=1&isAllowed=y

- Mottet A and Tempio G (2017). Global poultry production: Current state and future outlook and challenges. Worlds Poultry Science Journal, 73(2): 245-56. DOI: https://www.doi.org/10.1017/S0043933917000071
- Mousa-Balabel T, Al-Midany S, and Algazzar W (2021). Dim blue light colour reduces the activities and improves the performance of Indian river broilers under Egyptian conditions. Journal of the Hellenic Veterinary Medical Society, 72(3): 3171-3178. DOI: https://www.doi.org/10.12681/jhvms.28511
- Nawab A, Ibtisham F, Li G, Kieser B, Wu J, Liu W, Zhao Y, Nawab Y, Li K, Xiao M, et al. (2018). Heat stress in poultry production: Mitigation strategies to overcome the future challenges facing the global poultry industry. Journal of Thermal Biology, 78: 131-139. DOI: <u>https://www.doi.org/10.1016/j.jtherbio.2018.08.010</u>
- Nawaz AH, Amoah K, Leng QY, Zheng JH, Zhang WL, and Zhang L (2021). Poultry response to heat stress: Its physiological, metabolic, and genetic implications on meat production and quality including strategies to improve broiler production in a warming world. Frontiers in Veterinary Science, 8: 699081. DOI: https://www.doi.org/10.3389/fvets.2021.699081
- Nilsson JA, Molokwu MN, and Olsson O (2016). Body temperature regulation in hot environments. PLoS One, 11(8): e0161481. DOI: https://www.doi.org/10.1371/journal.pone.0161481
- Nochi T, Jansen CA, Toyomizu M, and van Eden W (2018). The welldeveloped mucosal immune systems of birds and mammals allow for similar approaches of mucosal vaccination in both types of animals. Frontiers in Nutrition, 5: 60. DOI: https://www.doi.org/10.3389/fnut.2018.00060
- Nyoni NMB, Grab S, and Archer ERM (2019). Heat stress and chickens: climate risk effects on rural poultry farming in low-income countries. Climate and Development, 11(1): 83-90. DOI: https://doi.org/10.1080/17565529.2018.1442792
- Oloyo A and Ojerinde A (2019). Poultry Housing and Management. In Poultry; IntechOpen., London, UK. Available at: https://cdn.intechopen.com/pdfs/65864.pdf
- Orakpoghenor O, Ogbuagu NE, and Sa'Idu L (2020). Effect of environmental temperature on water intake in poultry. In: A. Kumar Patra (Editor), Advances in poultry nutrition research, IntechOpen., pp. 1-8. DOI: <u>https://www.doi.org/10.5772/intechopen.95695</u>
- Pacheco LG, Sakomura NK, Suzuki RM, Dorigam JCP, Viana GS, Van Milgen J, and Denadai JC (2018). Methionine to cystine ratio in the total sulfur amino acid requirements and sulfur amino acid metabolism using labelled amino acid approach for broilers. BMC Veterinary Research, 14(1): 364. DOI: https://www.doi.org/10.1186/s12917-018-1677-8
- Park S, Park B, and Hwangbo J (2015). Effect of cold water and inverse lighting on growth performance of broiler chickens under extreme heat stress. Journal of Environmental Biology, 36(4): 865-873. Available

https://www.cabdirect.org/cabdirect/abstract/20153404891

- Park SO and Kim WK (2016). Effects of betaine on biological functions in meat-type ducks exposed to heat stress. Poultry Science, 96(5): 1212-1218. DOI: https://www.doi.org/10.3382/ps/pew359
- Parvin R, Mushtaq MMH, Kim MJ, and Choi HC (2014). Light emitting diode (LED) as a source of monochromatic light: A novel lighting approach for behaviour, physiology, and welfare of poultry. World's Poultry Science Journal, 70(3): 543-556. DOI: https://www.doi.org/doi:10.1017/S0043933914000609
- Pawar S, Sajjanar B, Lonkar V, Kurade N, Kadam A, Nirmal A, Brahmane M, and Bal S (2016). Assessing and mitigating the impact of heat stress on poultry. Advances in Animal and Veterinary Sciences, 4: 332-341. DOI: http://www.doi.org/10.14737/journal.aavs/2016/4.6.332.341
- Piestun Y, Patael T, Yahav S, Velleman SG, and Halevy O (2017). Early post-hatch thermal stress affects breast muscle development and satellite cell growth and characteristics in broilers. Poultry Science, 96(8): 2877-2888. DOI: <u>https://www.doi.org/10.3382/ps/pex065</u>

- Popoola IO, Popoola OR, Ojeniyi MO, Olajide OO, and Iyayi EA (2019). The roles of key electrolytes in balancing blood acid-base and nutrient in broiler chickens reared under tropical conditions. Natural Science, 12(1): 4-11. DOI: https://www.doi.org/10.4236/ns.2020.121002
- Pursey KM, Davis C, and Burrows TL (2017). Nutritional aspects of food addiction. Current Addiction Reports, 4: 142-150. DOI: https://www.doi.org/10.1007/s40429-017-0139-x
- Qaid MM and AlGaradi MA (2021). Protein and amino acid metabolism in poultry during and after heat stress: A review. Animals, 11(4): 1167. DOI: <u>https://www.doi.org/10.3390/ani11041167</u>
- Quilumba C, Quijia E, Gernat A, Murillo G, and Grimes J (2015). Evaluation of different water flow rates of nipple drinkers on broiler productivity. Journal of Applied Poultry Research, 24(1): 58-65. DOI: <u>https://www.doi.org/10.3382/japr/pfv005</u>
- Quinteiro-Filho WM, Calefi AS, Cruz DSG, Aloia TPA, Zager A, Astolfi-Ferreira CS, Piantino FJA, Sharif S, and Palermo-Neto J (2017). Heat stress decreases the expression of the cytokines, avian β-defensins 4 and 6 and Toll-like receptor 2 in broiler chickens infected with *Salmonella enteritidis*. Veterinary Immunology and Immunopathology, 186(1): 19-28. DOI: https://www.doi.org/10.1016/j.vetimm.2017.02.006
- Rahman H and Hidayat C (2020). Reducing the negative effect of heat stress in broiler through nutritional and feeding strategy. IOP Conference Series: Earth Environmental Science, 465: 012034. DOI: https://www.doi.org/10.1088/1755-1315/465/1/012034
- Rajkumar U, Reddy BLN, Rajaravindra K, Niranjan M, Tarun B, Chatterjee RN, Arun P, Reddy RM, and Sharma RP (2010). Effect of naked neck gene on immune competence, serum biochemical and carcass traits in chickens under a tropical climate. Asian-Australasian Journal of Animal Sciences, 23(7): 867-872. DOI: https://www.doi.org/10.5713/ajas.2010.90548
- Ranjan A, Sinha R, Devi I, Rahim A, and Tiwari S (2019). Effect of heat stress on poultry production and their management approaches. International Journal of Current Microbiology and Applied Sciences, 8(2): 1548-1555. DOI: https://www.doi.org/10.20546/ijcmas.2019.802.181
- Ratcliffe MJ and Härtle S (2014). B cells, the bursa of Fabricius and the generation of antibody repertoires. In: K. A. Schat, B. Kaspers, P. Kaiser (Editors), Avian immunology, 2nd Edition. Academic Press., San Diego. pp. 65-89.
- Rath KP, Behura CN, Sahoo PS, Panda P, Mandal DK, and Panigrahi NP (2015). Amelioration of heat stress for poultry welfare: A strategic approach. International Journal of Livestock Research, 5(3): 1-9. DOI: <u>https://www.doi.org/10.5455/JJLR.20150330093915</u>
- Rath M, Muller I, Kropf P, Closs EL, and Munder M (2014). Metabolism viaarginase or nitric oxide synthase: Two competing arginine pathways in macrophages. Frontiers in Immunology Inflammation, 5: 532. DOI: <u>https://www.doi.org/10.3389/fimmu.2014.00532</u>
- Reddy TE and Ramya P (2015). Heat stress strategies for layers in hot climates. Available at: <u>https://www.wattagnet.com/articles/22314-heat-stress-strategies-for-layers-in-hot-climates</u>
- Riber AB (2015). Effects of color of light on preferences, performance, and welfare in broilers. Poultry Science, 94(8): 1767-1775. DOI: https://www.doi.org/10.3382/ps/pev174
- Robert L, Francisco JT, Ulf T, and Ingrid H (2017). Nanoparticle-based hyperthermia distinctly impacts the production of ROS, expression of Ki-67, TOP2A, and TPX2, and induction of apoptosis in pancreatic cancer. International Journal of Nanomedicine, 12: 1009-1018. DOI: <u>http://www.doi.org/10.2147/IJN.S108577</u>
- Rostagno HM (2020). Effects of heat stress on the gut health of poultry. Journal of Animal Science, 98(4): skaa090. DOI: https://www.doi.org/10.1093/jas/skaa090
- Ruuskanen S, Hsu YB, and Nord A (2021). Endocrinology and thermoregulation of birds in changing climate. Molecular and Cellular Endocrinology, 519: 111088. DOI: <u>https://www.doi.org/10.1016/j.mce.2020.111088</u>
- Saeed M, Abbas G, Alagawany M, Kamboh AA, Abd El-Hack ME, Khafaga AF, and Chao S (2019). Heat stress management in poultry farms: A comprehensive overview. Journal of Thermal Biology, 84: 414-425. DOI: <u>https://www.doi.org/10.1016/j.jtherbio.2019.07.025</u>

- Saeed M, Babazadeh D, Naveed M, Arain MA, Hassan FU, and Chao S (2017). Reconsidering betaine as a natural anti-heat stress agent in poultry industry: a review. Tropical Animal Health and Production, 49(7): 1329-1338. Available at: https://europepmc.org/article/med/28733762
- Saelao P, Wang Y, Chanthavixay G, Yu V, Gallardo RA, Dekkers JCM, Lamont SJ, Kelly T, and Zhou H (2021). Distinct transcriptomic response to Newcastle disease virus infection during heat stress in chicken tracheal epithelial tissue. Scientific Reports, 11(1): 7450. DOI: <u>https://www.doi.org/10.1038/s41598-021-86795-x</u>
- Safdar AHA and Maghami SPMG (2014). Heat stress in poultry: Practical tips. European Journal of Experimental Biology, 4: 625-631.
- Sahin K, Orhan C, Tuzcu M, Sahin N, Hayirli A, Bilgili S, and Kucuk O (2016). Lycopene activates antioxidant enzymes and nuclear transcription factor systems in heat-stressed broilers. Poultry Science, 95(5): 1088-1095. DOI: https://www.doi.org/10.3382/ps/pew012
- Saleeva IP, Sklyar AV, Marinchenko TE, Postnova MV, Ivanov AV, and Tikhomirov AI (2019). Feasibility study on innovative energy-saving technologies in poultry farming. E3S Web of Conferences, 124: 05070. DOI: https://www.doi.org/10.1051/e3sconf/201912405070
- Saleh AA, Shurky M, Farrag F, Soliman MM, and Moneim AMEA (2021). Effect of feeding wet feed or wet feed fermented by *Bacillus licheniformis* on growth performance, histopathology growth and lipid metabolism marker genes in broiler chickens. Animals, 11(1): 83. DOI: <u>https://www.doi.org/10.3390/ani11010083</u>
- Santos RR, Awati A, Roubos-van den Hil PJ, Tersteeg-Zijderveld MHG, Koolmees PA, and Fink-Gremmels J (2015). Quantitative histomorphometric analysis of heat-stress-related damage in the small intestines of broiler chickens. Avian Pathology, 44(1): 19-22. DOI: <u>https://www.doi.org/10.1080/03079457.2014.988122</u>
- Scanes CG (2015). Sturkie's avian physiology. Regulation of body temperature: Strategies and mechanisms, 6th Edition. Academic Press., USA. Chapter 37, pp. 869-905.
- Seifi K, Rezaei M, Yansari AT, Zamiri MJ, Riazi GH, and Heidari R (2020). Short chain fatty acids may improve hepatic mitochondrial energy efficiency in heat stressed-broilers. Journal of Thermal Biology, 89: 102520. DOI: https://www.doi.org/10.1016/j.jtherbio.2020.102520
- Sejian V, Bhatta R, Gaughan JB, Dunshea FR, and Lacetera N (2018). Review: Adaptation of animals to heat stress. Animal, 12(2): 431-44. DOI: <u>https://www.doi.org/10.1017/S1751731118001945</u>
- Shakeri M and Le HH (2022). Deleterious effects of heat stress on poultry production: Unveiling the benefits of betaine and polyphenols. Poultry, 1(3): 147-156. DOI: https://www.doi.org/10.3390/poultry1030013
- Shakeri M, Oskoueian E, Le HH, and Shakeri M (2020). Strategies to combat heat stress in broiler chickens: Unveiling the roles of selenium, vitamin E and vitamin C. Veterinary Sciences, 7(2): 71. DOI: <u>https://www.doi.org/10.3390/vetsci7020071</u>
- Shehata AM, Saadeldin IM, Tukur HA, and Habashy WS (2020). Modulation of heat-shock proteins mediates chicken cell survival against thermal stress. Animals, 10(12): 2407. DOI: https://www.doi.org/10.3390/ani10122407
- Sinha B, Mandal K, Kumari R, and Kumari T (2018). Estimate and effect of breeds on egg quality traits of poultry - A review. International Journal of Livestock Research, 8: 8-21. DOI: <u>https://www.doi.org/10.5455/ijlr.20170812102444</u>
- Skřivan M, Milan M, Michaela E, and Skřivanová V (2013). Influence of dietary vitamin C and selenium, alone and in combination, on the performance of laying hens and quality of eggs. Czech Journal of Animal Science, 58: 91-97. DOI: http://www.doi.org/10.17221/6619-CJAS
- Slawinska A, Mendes S, Dunislawska A, Siwek M, Zampiga M, Sirri F, Meluzzi A, Tavaniello S, and Maiorano G (2019). Avian model to mitigate gut-derived immune response and oxidative stress during the heat. BioSystems, 178: 10-15. DOI: https://www.doi.org/10.1016/j.biosystems.2019.01.007
- Smalling S, Diarra SS, and Amosa F (2019). Effect of feed form and water addition on growth performance of finishing broilers in a hot

humid environment. Pakistan Journal of Nutrition, 18(4): 339-345. DOI: <u>https://www.doi.org/10.3923/pjn.2019.339.345</u>

- Smith BA, Meadows S, Meyers R, Parmley EJ, and Fazil A (2019). Seasonality and zoonotic foodborne pathogens in Canada: Relationships between climate and Campylobacter, E. coli and Salmonella in meat products. Epidemiology and Infection, 147: e190. DOI: https://www.doi.org/10.1017/S0950268819000797
- Sohsuebngarm D, Kongpechr S, and Sukon P (2019). Microclimate, body weight uniformity, body temperature, and footpad dermatitis in broiler chickens reared in commercial poultry houses in hot and humid tropical climates. World's Veterinary Journal, 9(4): 241-248. DOI: <u>https://www.doi.org/10.36380/scil.2019.wvj30</u>
- Song DJ and King AJ (2015). Effects of heat stress on broiler meat quality. Worlds Poultry Science Journal, 71(4): 701-709. DOI: https://www.doi.org/10.1017/S0043933915002421
- Souza LFA, de Espinha LP, de Almeida EA, Lunedo RA, Furlan RL, and Macari M (2016). How heat stress (continuous or cyclical) interferes with nutrient digestibility, energy and nitrogen balances and performance in broilers. Livestock Science, 192: 39-43. DOI: http://www.doi.org/10.1016/j.livsci.2016.08.014
- Suganya T, Senthilkumar S, Deepa K, and Amutha R (2015). Nutritional management to alleviate heat stress in broilers. International Journal of Science, Environment and Technology, 4(3): 661-666. Available at: https://www.ijset.net/journal/683.pdf
- Sugiharto S, Turrini Y, Isroli, I, Endang W, and Endang K (2017). Dietary supplementation of probiotics in poultry exposed to heat stress—A review. Annals for Animal Science, 17(3): 591-604. DOI: https://www.doi.org/10.1515/aoas-2016-0062
- Surai PF, Kochish II, Fisinin VI, and Kidd MT (2019). Antioxidant defence systems and oxidative stress in poultry biology: An update. Antioxidants, 8(7): 235. DOI: http://www.doi.org/10.3390/antiox8070235
- Syafwan S, Kwakkel R, and Verstegen M (2011). Heat Stress and feeding strategies in meat-type chickens. World's Poultry Science Journal, 67(4): 653-674. DOI: https://www.doi.org/10.1017/S0043933911000742
- Tamzil MH (2014). Heat stress on poultry: Metabolism, effects and efforts to overcome. Indonesian Bulletin of Animal and Veterinary Sciences, 24(2): 57-66. DOI: https://www.doi.org/10.14334/wartazoa.v24i2.1049
- Tang LP, Liu YL, Zhang JX, Ding KN, Lu MH, and He YM (2022). Heat stress in broilers of liver injury effects of heat stress on oxidative stress and autophagy in liver of broilers. Poultry Science, 101(10): 102085. DOI: <u>https://www.doi.org/10.1016/j.psj.2022.102085</u>
- Tari AA, Sadeghi AA, and Mousavi NS (2020). Dietary vegetable oils inclusion on the performance, hormonal levels and HSP 70 gene expression in broilers under heat stress. Acta Scientiarum Animal Sciences, 42(1): e45517 DOI: https://www.doi.org/10.4025/actascianimsci.v42i1.45517
- Teyssier JR, Brugaletta G, Sirri F, Dridi S, and Rochell SJ (2022). A review of heat stress in chickens. Part II: Insights into protein and energy utilization and feeding. Frontiers in Physiology, 13: 943612. DOI: https://www.doi.org/10.3389/fphys.2022.943612
- Tóth R, Szabadi TN, Lázár B, Buda K, Végi B, Barna J, Patakiné Várkonyi PE, Liptói K, Pain B, and Gócza E (2021). Effect of posthatch heat-treatment in heat-stressed Transylvanian naked neck chicken. Animals, 11(6): 1575. DOI: https://www.doi.org/10.3390/ani11061575
- Van Goor A, Ashwell CM, Persia ME, Rothschild MF, Schmidt CJ, and Lamont SJ (2017). Unique genetic responses were revealed in RNAseq of the spleen of chickens stimulated with lipopolysaccharide and short-term heat. PLoSOne, 12(2): e0171414. DOI: https://www.doi.org/10.1371/journal.pone.0171414
- Vandana GD and Sejian V (2018). Towards identifying climate-resilient poultry birds. Journal of Dairy, Veterinary and Animal Research, 7(3): 84-85. DOI: <u>http://www.doi.org/10.15406/jdvar.2018.07.00195</u>
- Vandana GD, Sejian V, Lees AM, Pragna P, Silpa MV, and Maloney SK (2021). Heat stress and poultry production: Impact and amelioration. International Journal of Biometeorology, 65(2): 163-179. DOI: <u>https://www.doi.org/10.1007/s00484-020-02023-7</u>

- Varasteh S, Braber S, Akbari P, Garssen J, and Fink-Gremmels J (2015). Differences in susceptibility to heat stress along the chicken intestine and the protective effects of galactooligosaccharides. PLoS One, 10: e0138975. DOI: <u>https://www.doi.org/10.1371/journal.pone.0138975</u>
- Ventura MVA and Matias da Silva R (2019). Bone problems are caused by the deficiency of calcium and phosphorus in the feeding of broilers. Biomedical Journal of Scientific and Technical Research, 16(4): 12223-12229. DOI: https://www.doi.org/10.26717/BJSTR.2019.16.002886
- Waiz AH, Gautam L, Nagada R, and Bhat AG (2016). Effect of wet feeding on feed conversion efficiency in laying hens during the summer season. Iranian Journal of Applied Animal Science, 6(2): 383-387. Available at: <u>https://ijas.rasht.iau.ir/article_522858.html</u>
- Wang JP, Lee JH, Jang HD, Yan L, Cho JH, and Kim IH (2011). Effects of delta-aminolevulinic acid and vitamin C supplementation on iron status, production performance, blood characteristics and egg quality of laying hens. Journal of Animal Physiology and Animal Nutrition, 95(4): 417-423. DOI: <u>http://www.doi.org/10.1111/j.1439-0396.2010.01067.x</u>
- Wang W, Yan F, Hu J, Amen O, and Cheng H (2018). Supplementation of Bacillus subtilis-based probiotic reduces heat stress-related behaviors and inflammatory response in broiler chickens. Journal of Animal Science, 96(5): 1654-1666. DOI: https://www.doi.org/10.1093/jas/sky092
- Wang Y, Saelao P, Chanthavixay K, Gallardo R, Bunn D, Lamont SJ, Dekkers JM, Kelly T, and Zhou H (2018). Physiological responses to heat stress in two genetically distinct chicken inbred lines. Poultry Science, 97(3): 770-780. DOI: https://www.doi.org/10.3382/ps/pex363
- Wang Y, WeiChao Z, HaiPeng S, Jiang T, and BaoMing L (2018). Roof insulation improving thermal environment and laying performance of poultry houses in summer. Transactions of the Chinese Society of Agricultural Engineering, 34(17): 207-213. Available at: https://www.cabdirect.org/cabdirect/abstract/20183344426
- Wasti S, Sah N, and Mishra B (2020). Impact of heat stress on poultry health and performances, and potential mitigation strategies. Animals, 10(8): 1266. DOI: https://www.doi.org/10.3390/ani10081266
- Wells KL, Hadad Y, Ben-Avraham D, Hillel J, Cahaner A, and Headon DJ (2012). Genome-wide SNP scan of pooled DNA reveals a nonsense mutation in FGF20 in the scaleless line of featherless chickens. BMC Genomics, 13: 257. DOI: <u>https://www.doi.org/10.1186/1471-2164-13-257</u>
- Wu Y, Huang J, Quan S, and Yang Y (2022). Light regimen on health and growth of broilers: an update review. Poultry Science, 101(1): 101545. DOI: <u>https://www.doi.org/10.1016/j.psj.2021.101545</u>
- Xie J, Tang L, Lu L, Zhang L, Lin X, Liu HC, Odle J, and Luo X (2015). Effects of acute and chronic heat stress on plasma metabolites,

hormones and oxidant status in restrictedly fed broiler breeders. Poultry Science, 94(7): 1635-1644. DOI: https://www.doi.org/10.3382/ps/pev105

- Xue B, Song J, Liu L, Luo J, Tian G, and Yang Y (2017). Effect of epigallocatechin gallate on growth performance and antioxidant capacity in heat-stressed broilers. Achieves of Animal Nutrition, 71(5): 362-372. DOI: https://www.doi.org/10.1080/1745039X.2017.1355129
- Yahav S (2015). Regulation of body temperature: Strategies and mechanisms. G. Colin. Scanes, (Editor), Sturkie's avian physiology. Academic Press., London, UK. pp. 869-905.
- Yousaf A, Jabbar A, Rajput N, Memon A, Shahnawaz R, Mukhtar N, Farooq F, Abbas M, and Khalil R (2019). Effect of environmental heat stress on performance and carcass yield of broiler chicks. World's Veterinary Journal, 9(1): 26-30. DOI: https://www.doi.org/10.36380/scil.2019.wyj4
- Zaboli G, Huang X, Feng X, and Ahn DU (2019). How can heat stress affect chicken meat quality? - A review. Poultry Science, 98(3): 1551-1556. DOI: <u>https://www.doi.org/10.3382/ps/pey399</u>
- Zaboli GR, Rahimi S, Shariatmadari F, Torshizi MAK, Baghbanzadeh A, and Mehri M (2017). Thermal manipulation during pre and post hatch on thermo-tolerance of male broiler chickens exposed to chronic heat stress. Poultry Science, 96(2): 478-485. DOI: https://www.doi.org/10.3382/ps/pew344
- Zarghi H, Golian A, and Yazdi FT (2020). Effect of dietary sulphur amino acid levels and guanidinoacetic acid supplementation on performance, carcase yield and energetic molecular metabolites in broiler chickens fed wheat-soy diets. Italian Journal of Animal Science, 19(1): 951-959. DOI: https://www.doi.org/10.1080/1828051X.2020.1809537
- Zeferino CP, Komiyama CM, Pelícia VC, Fascina VB, Aoyagi MM, Coutinho LL, Sartori JR, and Moura ASAMT (2016). Carcass and meat quality traits of chickens fed diets concurrently supplemented with vitamins C and E under constant heat stress. Animal, 10(1): 163-171. DOI: https://www.doi.org/10.1017/s1751731115001998
- Zerjal T, Gourichon D, Rivet B, and André B (2013). Performance comparison of laying hens segregating for the frizzle gene under thermoneutral and high ambient temperatures. Poultry science, 92(6): 1474-1485. DOI: <u>https://www.doi.org/10.3382/ps.2012-02840</u>
- Zhang C, Zhao XH, Yang L, Chen XY, Jiang RS, Jin SH, and Geng ZY (2017). Resveratrol alleviates heat stress-induced impairment of intestinal morphology, microflora, and barrier integrity in broilers. Poultry Science, 96(12): 4325-4332. DOI: https://www.doi.org/10.3382/ps/pex266
- Zhang X, Owens CM, and Schilling MW (2017). Meat: the The edible flesh from mammals only or does it include poultry, fish, and seafood?. Animal Frontiers, 7(4): 12-8. DOI: https://www.doi.org/10.2527/af.2017.0437